Synchronous primary sigmoid colon cancer and primary thyroid cancer followed by a malignant tumor of the kidney: Case report of multiple primary cancer and review of the literature

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Abstract. Multiple primary cancer (MPC) is relatively rare. With the development of diagnostic and anti-cancer therapeutic techniques, the incidence of MPC is rising annually. However, the incidence of triple or quadruple cancers in a single patient remains low. In this report, the case of a 58-year-old male with triple malignant cancer is outlined. Synchronous sigmoid colon cancer and thyroid cancer were diagnosed in May 2015; on subsequent re-examination, metastasis to the liver and a malignant kidney tumor were also identified. The diagnosis was established via computed tomography (CT), Positron emission tomography-CT (PET-CT) and other laboratory examination results, including analysis of tumor markers and liver function, and was confirmed by pathological diagnosis. The patient underwent radical surgery and standardized chemotherapy. Through literature review, the definition, characteristics, classification, incidence, possible causes of and treatment strategies for MPC were more clearly understood. In addition, immunohistochemical staining of integrin αvβ6 was performed on patient tissue specimens, where integrin αvβ6 expression was confirmed in cancer of the colon, thyroid and liver, as a result of colonic metastasis. Therefore, the involvement of integrin αvβ6 in the malignant progression of MPC was hypothesized, which may aid the investigation of MPC etiology in the future.

Case report

In October 2016, a 58-year-old male patient was referred to the Hepatobiliary Department of Qilu Hospital (Jinan, China) for review. The patient had previously been diagnosed with sigmoid colon cancer (first primary) and left lobe thyroid cancer (second primary) in May 2015. A left hemicolectomy and radical surgery were undertaken for the thyroid cancer. The final pathology results for sigmoid colon cancer revealed well-differentiated adenocarcinoma, polypus type, incisal surface square 2.8x0.3 cm, and invasion of the stratum sub-mucosa. There were no carcinoma cells at the anodic and bottom incisal margin, and no metastasis to the lymph nodes (Fig. 1A). The final pathology results for thyroid cancer revealed left lobe micro-papillary carcinoma, and right lobe lymphatic nodule thyroiditis (Fig. 1B). As the sigmoid colon cancer was in the early stages (pT1, pN0, M0; Stage I), adjuvant chemotherapy was not administered; 3x cantharidis capsules were taken twice daily, for 11 months. Routine, post-surgical, examination, including blood tests, abdominal and pelvic CT scans, and a colonoscopy were performed. Tumor marker analysis indicated that carcinoembryonic antigen (CEA) (230.10 ng/ml), cancer antigen (CA)-199 (89.23 U/ml) and CA-724 (8.80 U/ml), and neuron-specific enolase (NSE) (21.68 ng/ml), among other markers, including CYFr21-1, squamous cell carcinoma antigen, pro-gastrin releasing peptide, α-fetoprotien, CA-125, Ferrari, total prostate-specific antigen and serum sialic acid were within the normal range. CT scan suggested that, according to the patient history, the space-occupying lesion located in the V and VIII liver segments may have been the metastatic tumor; the VII segment tumor may have been an hemangioma (Fig. 2A and B); and the space-occupying lesion of the right kidney anus perineum may have been primary kidney cancer (third primary) (Fig. 2C and D).

The colonoscopy results were satisfactory, with no recurrence noted (Fig. 2E). To further investigate the characteristics of the liver and kidney tumors, PET-CT examination was conducted, the results of which indicated that: i) The tumor in the right anterior lobe of the liver may have been a metastatic lesion (Fig. 3A); ii) combined with the CT scan results, it was supposed that the tumor of the right kidney anus perineum may have been malignant (hypo-metabolism type) (Fig. 3B); iii) 1.5 years after radical surgery for left lobe thyroid cancer,
no recurrence was observed (Fig 3C); iv) a small node in the left anterior-inferior abdominal exhibited high absorbance of FDG, possibly indicating metastasis or a responsive modification (Fig. 3D).

As a result of these findings, surgical excision of the metastatic liver tumor and hepatic hemangiomia, in addition to a partial nephrectomy, were conducted on October 11th, 2016 (Fig. 4). The final pathology examination results revealed: i) V and VIII segment liver, well-differentiated adenocarcinoma in the liver tissue (incision square, 4.0x2.7 cm), which, according to the patient history, is the metastatic tumor from the primary colon cancer (Fig. 1C); ii) VII segment liver, cavernous hemangiomia; iii) kidney, clear-cell carcinoma of the kidney (II level nuclear) with a diameter of 2.3 cm (Fig. 1D); iv) abdominal wall, hyperplastic collagen fiber and vascular tissue. Post-surgery, the patient recovered well, and was administered one course of FOLFORx6 chemotherapy prior to discharge (November 3rd, 2016). In subsequent follow-up, the patient had recovered markedly.

For resected specimens of colon, thyroid, liver metastatic tumor and renal cell carcinoma, immunohistochemical staining of integrin αvβ6 was performed, the results of which are detailed below. Experimental methods were as follows: Mouse anti-human monoclonal antibody (clone 6.2A1; IgG1) against integrin αvβ6 was obtained from Biogen (Abcam, Cambridge, MA, USA). Patient tissue sections were depaaffinized and hydrated, and heat-induced epitope retrieval was performed using Borg decloaking high pH buffer in the Biocare decloaking chamber (Biocare Medical, LLC, Pacheco, CA, USA). Endogenous peroxidase activity was blocked with 3% hydrogen peroxide for 5 min at room temperature. The slides were incubated with an Endogenous Avidin-Biotin Blocking kit (cat. no. ab64212; Abcam) for 10 min, followed by further incubations with the integrin αvβ6 primary antibody 6.2A1 (1:500) overnight at 4˚C. The following day, biotinylated anti-mouse IgG (1:200; cat. no. ab207996; Abcam) was applied to the slides for 20 min at 37˚C, which were subsequently treated with 50 µl horseradish peroxidase-labeled streptavidotibiotin (Dako; Agilent Technologies, Inc., Santa Clara, CA, USA) for 15 min at 37˚C and Betazoid Diaminobenzidine (Biocare Medical, LLC, Pacheco, CA, USA) for color development for 2 min at room temperature. The slides were counterstained with hematoxylin for 1 min (Dako; Agilent Technologies, Inc.), rinsed with water for 2 min, and dehydrated with alcohol and xylene for several seconds, before a coverslip was affixed. Normal mouse IgG (1:200; cat. no. ab188776; Abcam) was substituted for primary antibody as the negative control and incubated with the slides overnight at 4˚C. Unless otherwise indicated, incubations were performed at room temperature. On all slides, >5% of the cells stained positively for integrin αvβ6.

Discussion

MPC refers to the presence of ≥2 pathologically diagnosed primary cancers, in ≥1 organ of the same patient. The diagnosis of MPC is based on the Warren standards (1): i) All tumors identified after the pathological examination are malignant; ii) each tumor is independent of the others, and the possibility of transfer is excluded; and iii) all tumors originate from different organs. According to differences in primary tumor occurrence times, simultaneous MPC refers to cases wherein all tumors appear within 6 months; otherwise, it is known as non-simultaneous or metachronous MPC.

MPC is relatively rare, with a reported incidence range of 0.8-6.6% among the general population (2-4); however, the Surveillance Epidemiology and End Results report issued by the National Cancer Institute states that between 1975 and 2000, the incidence increased to ~16% (5). This difference may be associated with location, environment, living habits, quality and availability of medical care, the type of cancer and statistical research methods. The incidence of triple or quadruple malignant cancer in one patient remains rare, with the incidence of quadruple cancer reported to be <0.1% (6-8). Thus, cases of MPC may be overlooked or misdiagnosed. MPC typically originates in pairs of organs or same system organs, such as the gastrointestinal, respiratory or urological systems. Male patients have an increased tendency to develop gastrointestinal cancer, and female patients are more prone to breast and gastrointestinal cancer (9,10). Therefore, when a patient presents with these particular cancer types, clinicians should investigate whether an accompanying malignancy has occurred elsewhere. Baigrie (11) reported that colon cancer is frequently present in cases of MPC, particularly in patients between 50 and 60 years of age.

The definite cause of MPC is not clear. Though there are a number of factors that may influence its occurrence (including smoking, and exposure to carcinogens or carcinogenic environments) there are numerous defining characteristics of MPC. With improvements in living standards and medical technology, life expectancy is increasing, in accordance with the probability of acquiring a malignancy. It was reported by Curtis et al (12) that patients with cancer were 14% more likely to develop an additional tumor compared with the general population. Additionally, it was deduced by Jiao et al (6) that this phenomenon was associated with increased genetic instability, and the reduction of tumor immunity in cancer patients.

In addition, the wide application of diagnostic imaging has led to increased radiation exposure; this is particularly prominent in developed countries (13), where during routine follow-up examinations, patients are exposed through imaging procedures including X-rays and CT scans (14). Frequent doses of radiation may increase the occurrence of further malignancies (15). Studies have confirmed that the occurrence of head and neck cancer, particularly thyroid cancer, is associated with long-term radiological examination (16); specifically, the over-diagnosis and treatment of small adenocarcinoma was reported to influence the occurrence of thyroid cancer (17,18). This is consistent with the case reported in the current study. Furthermore, the carcinogenic effects of chemotherapy and endocrine therapy may contribute to the development of MPC. For example, studies have demonstrated that in patients with breast cancer who received long-term treatment with tamoxifen or raloxifene, the incidence of endometrial cancer was increased 2-3-fold compared with healthy volunteers (19,20).

Integrin αvβ6 is an integrin subtype expressed only in epithelial cells; its primary ligand is fibronectin (FN). In healthy epithelial cells the expression of integrin αvβ6 is rare (21), although it is increased substantially in response to injury and/or inflammation, and in epithelial tumors (including
gastric carcinoma and colon cancer) (22-24). The *de novo* expression of integrin αvβ6 has been reported to modulate a number of characteristics of colon carcinoma cells, including adhesion and spreading on fibronectin, proliferation in collagen gels, tumor growth, invasion and metastasis, and apoptosis (25-28). In the present case report, immunohistochemical staining of integrin αvβ6 was conducted, which revealed positive expression of integrin αvβ6 in numerous tissues, including those from the colon cancer primary site (Fig. 1E), the left lobe of the thyroid (Fig. 1F), and the hepatic metastatic tumor (Fig. 1G). Expression of integrin αvβ6 was negative in clear cell carcinoma of the kidney (Fig. 1H).

Figure 1. Tissue slice photomicrographs. (A) Hematoxylin and eosin stain photomicrograph of sigmoid colon cancer (magnification, x100). (B) Hematoxylin and eosin stain photomicrograph of left lobe thyroid micro-papillary carcinoma (magnification, x100). (C) Hematoxylin and eosin stain photomicrograph of differentiated adenocarcinoma in liver tissue (magnification, x200). (D) Hematoxylin and eosin stain photomicrograph of right kidney clear-cell carcinoma (magnification, x200). (E) Immunohistochemistry photomicrographs of positive integrin αvβ6 staining in colon cancer tissue (magnification, x100). (F) Immunohistochemistry photomicrographs of positive integrin αvβ6 staining in thyroid micro-papillary carcinoma tissue (magnification, x100). (G) Immunohistochemistry photomicrographs of positive integrin αvβ6 staining in the hepatic metastatic tumor tissue (magnification, x200). (H) Immunohistochemistry photomicrographs of negative integrin αvβ6 staining in kidney clear-cell carcinoma tissue (magnification, x400). Integrin αvβ6 staining was observed predominantly in the cell membrane. Positive αvβ6 stained cells appeared with a brown appearance and negative stained cells appeared with a blue appearance, which was induced by hematoxylin stain.
was identified that the rate of positive integrin $\alpha_v\beta_6$ expression in hepatic metastatic foci was 71.4% (29). Our previous study regarding the associations between integrin $\alpha_v\beta_6$ and thyroid cancer revealed the positive expression rate of integrin

Figure 2. Abdominal CT scan. Arrowhead refers to the location of the lesion. (A) Plain CT scan of liver hemangioma; (B) arterial phase CT scan of liver hemangioma; (C) plain CT scan of right kidney lesion; (D) arterial phase CT scan of right kidney lesion; (E) colonoscopy examination images. CT, computed tomography.

Figure 3. Whole body positron emission tomography-computed tomography scan. Arrowhead refers to the location of the lesion. (A) Hypo-metabolism lesion located in the right anterior lobe of the liver; (B) hypo metabolism lesion located in the right kidney anus perineum; (C) normal metabolism in the neck region with no recurrence of previous thyroid cancer; (D) slight hypo-metabolism of lesion located in the left anterior-inferior abdominal wall.
\( \alpha \nu 6 \) in thyroid papillary carcinoma to be 79.03% (49/62), 78.57% (22/26) in thyroid follicular carcinoma, and 100% in metastatic lymph nodes (10/10)(30). Combined with additional previous data, it was proposed that integrin \( \alpha \nu 6 \) may have an important role in MPC.

Renal clear cell carcinoma (RCC) originates from the malignant transformation of renal tubular epithelial cells. Due to its expression in malignant epithelial tumors, integrin \( \alpha \nu 6 \) may also be expressed in RCC. However, in this patient study, RCC specimens were negative for integrin \( \alpha \nu 6 \) expression, supported by the lack of literature reporting integrin \( \alpha \nu 6 \) expression in RCC tissue. Negative expression is predicted to be due to the loss of cytoplasm from RCC carcinoma cells during the preparation of the tissue sections. The loss of lipid components and vacuolization of the cytoplasm forms the so-called ‘transparent’ component; as integrin \( \alpha \nu 6 \) is predominantly expressed in the epithelial cell membrane and cytoplasm, cytoplasmic depletion in RCC may potentially negate the expression of integrin \( \alpha \nu 6 \).

The incidence of MPC is increasing, thus there is an urgent need to develop the understanding of MPC pathogenesis, diagnosis and treatment. Current treatment regimens for MPC differ from those indicated for traditional recurrent or metastatic malignant tumors. For MPC, radical excision of the tumors is preferred. When faced with that for recurrent or metastatic malignant tumors, the prognosis for MPC is comparatively more favorable (31,32).

With improvements in anti-tumor treatment technology, the prognosis for tumor patients has improved, though there has been a corresponding rise in the incidence of MPC. The aim of the clinician should be to improve understanding of MPC, reduce misdiagnosis and missed diagnosis, make early diagnoses to facilitate earlier treatment, and to administer radical surgery, radiotherapy and chemotherapy, and targeted therapy to improve clearance rates and prolong patient survival. The current case demonstrates that integrin \( \alpha \nu 6 \) is positively expressed in MPC. Combined with previous research, this may implicate integrin \( \alpha \nu 6 \) in the pathogenesis of MPC, and aid future research into the etiology of MPC.

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Availability of data and materials
The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Authors' contributions
CP led patient diagnosis and treatment, and manuscript authorship. ZL performed the immunohistochemical staining. HG, was the resident carer, responsible for the daily care of the patient, recorded the clinical laboratory examination results, and retrieved the related literature. XZ was responsible for pathological procedures and diagnoses. CZ performed the radical nephrectomy. JN as the chief surgeon, performed the radical resection of sigmoid colon cancer and excision of liver metastases from sigmoid colon cancer. He was additionally involved in the immunohistochemical staining experiment and statistical analysis of data. As the Dean of the Institute of Laparoscopic Minimally Invasive Surgery of Shandong University, he gave the final approval of the article.

Ethics approval and consent to participate
Not applicable.

Patient consent for publication
The patient provided informed consent for the use of the data in this study.

Competing interests
The authors declare that they have no competing interests.

References