Abstract. The present study was designed to clarify the expression and prognostic significance of activated Akt and mTOR in cervical cancer and their correlation with response to neoadjuvant chemotherapy (NAC). Immunohistochemical analysis for p-Akt and p-mTOR expression was performed on paraffin-embedded biopsy specimens from 25 patients with advanced cervical cancer (stage Ib2-IIb). We correlated this finding with various clinicopathological variables and prognosis by uni- and multivariate analyses. All patients received cisplatin-based NAC, and primary tumor response was evaluated by RECIST criteria and then classified as a positive or negative response. Activation of Akt was detected in the cytoplasm and nucleus of the cancer cells in 12 patients (48%), whereas p-mTOR was detected in the cytoplasm and membrane of the cancer cells in 13 patients (52%). Post NAC evaluation of the primary tumor revealed 68% (17/25) responsive tumors. The expression of p-mTOR and distant metastasis significantly correlated with the response to NAC (p=0.0101 and p=0.0107); however, there was no significant correlation between p-Akt and p-mTOR expression and any of the clinicopathological characteristics of the patients. In the univariate analysis, activated Akt and mTOR were found to be significant prognostic indicators (p<0.05). In multivariate analysis, p-mTOR expression retained its significance as an independent poor prognostic marker (p=0.0178). In summary, our present study showed that cervical cancer expressed Akt and mTOR activation. Moreover, the expression of phosphorylated mTOR may have a role as a marker to predict response to chemotherapy and survival of cervical cancer patients who are treated with cisplatin-based neoadjuvant chemotherapy. Our results suggest that the mTOR cascade may be a promising target for therapeutic intervention in cervical cancer.

Introduction
Carcinoma of the cervix is the second most common neoplasm in women worldwide. Its incidence is the highest in developing countries, where organized screening programs with the Pap test have not been well implemented (1,2). The best possible treatment of locally advanced cervical cancer is a combination of radiation and cisplatin-based chemotherapy. However, the five-year overall survival is still only 52% (3), and the FIGO data from 1993 to 1995 also indicate that the prognosis of advanced cervical cancer remains poor (4). From this data, it is suggested that conventional treatment methods have reached a plateau and therefore, finding a good prognostic factor and predictor response to chemotherapy might be useful for overcoming this problem.

Akt or protein kinase B (PKB) is the cellular homologue product of the v-akt oncogene and is activated downstream from phosphatidylinositol 3-kinase (PI3K) by a variety of growth factors (5). Activation of Akt pathway has been proposed to be involved in oncogenesis and resistance to cancer therapy. The main consequences of Akt activation can be catalogued into three categories: survival, proliferation (increased cell number), and growth (increased cell size). One protein that is emerging as a central regulator of cell growth is mTOR, also known as FRAP1 (FKBP 12-rapamycin-associated protein 1), a serine/threonine kinase that serves as a molecular sensor in regulating protein synthesis on the basis of the availability of nutrients (6,7). However, the PI3K-Akt pathway is unlikely to be the only stimulus that leads to mTOR activation in cancer cells. For example, mTOR can also function as an ATP sensor (8). Recently, Wlodarski and colleagues reported that the activation of mTOR in transformed B lymphocytes is nutrient-dependent and independent of Akt, mitogen-activated protein kinase kinase, insulin growth factor-I, and serum (9). Therefore, the exact mechanism of the activation of mTOR signaling is still poorly understood.
This study was designed to observe the expression and prognostic significance of p-Akt and p-mTOR and their predictive value in the response to chemotherapy. Although several studies have reported that the expression of activated Akt and mTOR is implicated in human cancers, until the present time, to our knowledge, no reports on cervical cancer have been published.

Patients and methods

Patients. Archival biopsy specimens from 25 patients with advanced cervical cancer (stage Ib2-IIb) at the Gunma University Hospital, Japan, between 1995 and 2002 were analyzed and the study was conducted according to the ethical guidelines of our university. The ages of the patients ranged from 25 to 57 years, with a mean age of 45 years. Squamous cell carcinoma was the most frequent cancer subtype with 19 cases and there were 6 cases of adenosquamous carcinoma. Stages were determined according to the clinical criteria established by the International Federation of Gynecology and Obstetrics (FIGO) in 1994 (10). All patients had cisplatin-based NAC (POMP; cisplatin, vincristine, mitomycin-c). After 2 to 3 cycles of chemotherapy, it was decided whether radical hysterectomy was possible. Response evaluation to NAC was carried out every 3 months during follow-up based on RECIST (response evaluation criteria in solid tumors) criteria (11). The response was assessed with MRI. Patients were then divided into two groups, positive response (complete and partial response) and negative response (stable disease and progressive disease). Formalin-fixed and paraffin-embedded biopsies samples of the patients were obtained before the start of NAC and retrieved from the department of pathology after review of histological diagnosis on hematoxylin and eosin (H&E)-stained sections.

Immunohistochemistry. Immunohistochemical staining of the section for p-Akt and p-mTOR were performed using the streptavidin-biotin method. Sections (4-μm thick) were deparaffinized with xylene, rehydrated, and incubated with fresh 0.3% hydrogen peroxide in methanol for 30 min at room temperature. After rehydration through a graded ethanol series, the specimens were washed in phosphate-buffered saline (PBS). After a blocking treatment, the specimens were then incubated with the primary anti-p-Akt and anti-p-mTOR antibody (Cell Signaling Technology, Beverly, MA) at a dilution of 1:200, respectively in PBS containing 1% bovine serum albumin (BSA) at 4°C overnight. They were then washed with PBS and incubated in secondary antibody for 30 min at room temperature. Immunohistochemistry was performed using a Histofine SAB-PO (M) kit (Nichirei, Tokyo, Japan). The chromogen was a 3.3-0.02% solution containing 0.005% H2O2 in a 50 mM ammonium acetate-citric acid buffer, pH 6.0. The specimens were lightly counterstained with haematoxylin. Negative controls were prepared by omitting each primary antibody; no detectable staining was evident.

Evaluation. The staining evaluation was performed by two independent observers who did not have any knowledge of the clinical outcome. For p-Akt and p-mTOR, total staining was scored as the product of the staining intensity (on a scale of 0-3) x percentage of cells stained, resulting in a scale of 0-300. Staining intensity was scored as follows: 0, no appreciable staining in tumor cells; 1, barely detectable staining in cytoplasm, nucleus, or membrane, as compared with stromal elements; 2, readily appreciable brown staining distinctly marking tumor cell cytoplasm, nucleus, or membrane; and 3, dark brown staining in tumor cells completely obscuring cytoplasm, nucleus, or membrane (12). The data were then classified as null (0), weak (1-100), moderate (101-200), or strong (201-300) p-Akt or p-mTOR staining. This scoring system has been reported previously (13).

For purposes of statistical analysis, all cases staining at a null or weak level were grouped as negative and all cases staining at a moderate or strong level were grouped as positive.

Statistics. Statistical analysis was performed using the Stat View software program (version 5, SAS Institute, NC, USA). The unpaired two-group t-test was used for age, p-Akt expression and p-mTOR expression. A Chi-squared test was used for histological type, FIGO stage, lymphatic invasion, lymph node metastasis, distant metastasis, and venous invasion. Survival curves of the patients were calculated using the Kaplan-Meier method and analysis was performed using the log-rank test. The Cox proportional hazards model for the risk ratio was used to assess the simultaneous contribution of levels of p-Akt and p-mTOR to the survival rate. Significant differences were noted when P<0.05.

Results

Expression of p-Akt and p-mTOR in cervical cancer. To elucidate the expression status and localization of p-Akt and p-mTOR in cervical cancer patients, we performed immunohistochemical analysis, using p-Akt (Ser473) and p-mTOR (Ser2448) antibody on 25 biopsy samples from patients with advanced cervical cancer; paired normal cervical tissues were used as controls. Fig. 1 shows representative immunohistochemical staining of cervical cancer. Fig. 1A and E shows a negative control, in which the primary anti-p-Akt and p-mTOR was omitted. Negative controls were uniformly negative for immunostaining. Weak-to-strong nuclear and/or cytoplasmic staining of p-Akt was observed in primary tumor cells, and faint expression was seen in the surrounding stromal area (Fig. 1B-D). Positive staining of activated mTOR was detected in the cytoplasm and membrane (Fig. 1F-H). Positive staining for p-Akt and p-mTOR was seen in 12 of 25 (48%) and 13 of 25 (52%) biopsy specimens, respectively.

Correlation between clinicopathological outcome and the expression of p-Akt and p-mTOR. The correlations between the clinicopathological characteristics of patients with cervical cancer and the expression of p-Akt and p-mTOR in their tumors are summarized in Table I. Although cancer with activated Akt and mTOR tended to be associated with poorer lymphatic invasion status (p=0.13 and p=0.06), there was no significant correlation between positive p-Akt and p-mTOR with any cervical cancer pathological (histological type, lymphatic invasion, lymph node metastasis, venous invasion) or clinical (patient age and staging) outcome. The relationship between clinicopathological features and response
to NAC is summarized in Table II. The positive response of the primary tumor was 68% (17/25). We found that 91.67% (11/12) of the p-mTOR-negative group had a positive response, in comparison to 46.15% (6/13) of the p-mTOR-positive group. A statistical significance was observed between distant metastasis and p-mTOR expression and response to NAC (p=0.010 and p=0.011).

**Correlation between expression of p-Akt and p-mTOR and survival.** The expression of p-Akt and p-mTOR has been reported to be a predictor of poor prognosis in some cancers (13,14). The survival rates of patients with positive and negative expression of p-Akt and p-mTOR were then further analyzed. The survival rate for cervical cancer patients with negative p-Akt was significantly higher than that for patients with positive p-Akt expression (p=0.005; Fig. 2). The five-year survival rates for patients with negative and positive p-Akt were 52% and 48%, respectively. In addition, p-mTOR was also significantly correlated with the five-year survival rates (p=0.019; Fig. 3). To clarify whether their expression is a significant prognostic factor, uni- and multivariate survival analyses were performed, and the analyses showed that p-mTOR expression was independently associated with the prognosis of cervical cancer patients (Table III).

**Discussion**

The role of the activation of Akt and mTOR has been well characterized in many human cancers. Akt has a critical role in several signaling pathways, including survival, apoptosis, and proliferation. The well-known downstream target of Akt in survival signaling is mTOR (6).

This study showed that Akt and mTOR are expressed and activated in cervical cancer. Of the 25 cases of cervical cancer studied, positive activated Akt and mTOR expression was observed in 48% and 52%, respectively. Although PTEN (phosphatase and tensin homologue deleted on chromosome 10) is well characterized to negatively regulate PKB/Akt activation, [primarily through phosphatidylinositol-3,4,5-triphosphate (PIP3) dephosphorylation (15)], none of the cervical cancer cases showed mutation of the PTEN protein (16,17). As one possible explanation for our results, a previous study reported that the PIK3CA gene, which encodes the p110α catalytic subunit of PI3K, is frequently amplified in cervical cancer (18). In addition, a number of studies have discovered PKB/Akt gene amplifications in human cancers. No modified or mutated Akt genes have been found in mammals. Its activation may also be due to the activation of up-stream regulators of
PI3K, such as tyrosine-kinase, G-protein-coupled receptors, and integrins (19).

Similarly to previous reports (20-22), our present study showed that activated Akt was detected in the cytoplasm and nucleus of cervical cancer cells, whereas mTOR was observed in the cytoplasm and membrane. Other studies reported that p-Akt staining in breast and prostate cancer was mainly localized in the cell membrane (14,23) while, in epithelial ovarian carcinoma, a predominant nuclear localization was observed (24). It is well characterized that Akt is activated

Table II. Correlation of clinicopathological outcome with response to chemotherapy.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Total n=25</th>
<th>Response (+) n=17</th>
<th>Response (-) n=8</th>
<th>p value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (mean ± SD years)</td>
<td>45.0±9.0</td>
<td>46.710±7.573</td>
<td>43.875±10.176</td>
<td>0.443</td>
</tr>
<tr>
<td>Histological type</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Squamous</td>
<td>19</td>
<td>13</td>
<td>6</td>
<td>0.936</td>
</tr>
<tr>
<td>Adenosquamous</td>
<td>6</td>
<td>4</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>FIGO stage</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Ib2</td>
<td>3</td>
<td>3</td>
<td>0</td>
<td>0.445</td>
</tr>
<tr>
<td>Ila</td>
<td>3</td>
<td>2</td>
<td>1</td>
<td></td>
</tr>
<tr>
<td>IIb</td>
<td>19</td>
<td>12</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>Lympathic invasion</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>13</td>
<td>9</td>
<td>6</td>
<td>0.285</td>
</tr>
<tr>
<td>Present</td>
<td>12</td>
<td>8</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>Lymph node metastasis</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>17</td>
<td>12</td>
<td>5</td>
<td>0.686</td>
</tr>
<tr>
<td>Present</td>
<td>8</td>
<td>5</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>Distant metastasis</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>20</td>
<td>16</td>
<td>4</td>
<td>0.010*</td>
</tr>
<tr>
<td>Present</td>
<td>5</td>
<td>1</td>
<td>4</td>
<td></td>
</tr>
<tr>
<td>Venous invasion</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Absent</td>
<td>19</td>
<td>13</td>
<td>6</td>
<td>0.936</td>
</tr>
<tr>
<td>Present</td>
<td>6</td>
<td>4</td>
<td>2</td>
<td></td>
</tr>
<tr>
<td>p-Akt negative</td>
<td>13</td>
<td>11</td>
<td>2</td>
<td>0.059</td>
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<tr>
<td>Positive</td>
<td>12</td>
<td>6</td>
<td>6</td>
<td></td>
</tr>
<tr>
<td>p-mTOR negative</td>
<td>12</td>
<td>11</td>
<td>1</td>
<td>0.011*</td>
</tr>
<tr>
<td>p-mTOR positive</td>
<td>13</td>
<td>6</td>
<td>7</td>
<td></td>
</tr>
</tbody>
</table>

SD, standard deviation; FIGO, Federation Internationale de Gynecologie et d'Obstetrique; *Significant.

Table III. Prognostic factors of patients with invasive cervical cancer in Cox proportional hazards model.

<table>
<thead>
<tr>
<th>Risk factor</th>
<th>Univariate</th>
<th>Multivariate</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Hazard ratio</td>
<td>95% CI</td>
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<tr>
<td>Lymphatic invasion</td>
<td>1.436</td>
<td>0.649-3.448</td>
</tr>
<tr>
<td>Venous invasion</td>
<td>1.254</td>
<td>0.463-3.397</td>
</tr>
<tr>
<td>Lymph node metastasis</td>
<td>1.078</td>
<td>0.445-2.612</td>
</tr>
<tr>
<td>Distant metastasis</td>
<td>0.269</td>
<td>0.087-0.828</td>
</tr>
<tr>
<td>p-Akt expression in IHC</td>
<td>0.303</td>
<td>0.116-0.787</td>
</tr>
<tr>
<td>p-mTOR expression in IHC</td>
<td>0.042</td>
<td>0.182-0.990</td>
</tr>
</tbody>
</table>

IHC, immunohistochemistry; CI, confidence interval; *Significant.
in cells exposed to diverse stimuli, such as hormones, growth factors, and extracellular matrix components. The activation mechanism remains to be fully characterized, but it appears to occur downstream of PI3K. PI3K generates PIP3, a lipid second messenger essential for the translocation of Akt to the plasma membrane, where it is phosphorylated and activated by phosphoinositide-dependent kinase-1 (PDK1) and, possibly, other kinases (19). Akt directly phosphorylates and modifies its downstream effectors, such as mTOR, forkhead transcription factors FKHRL1, FHH1, and AFX, IKKα,
has been investigated and found. Activated mTOR in correlation with chemosensitivity and phosphorylated mTOR and chemotherapy response was evaluated based on RECIST criteria and mitomycin-c (POMP) before surgery, and the response of patients in this study were treated with cisplatin, vincristine, etoposide, and other drugs. We also investigated whether the activation of Akt and mTOR correlates with the response to NAC. The majority of patients in this study were treated with cisplatin-based neoadjuvant chemotherapy.

In conclusion, our present study showed that cervical cancer expressed Akt and mTOR activation and the activation of mTOR may have a potential role in predicting the response to chemotherapy and prognosis of advanced cervical cancer treated with cisplatin-based neoadjuvant chemotherapy. Therefore, mTOR might be a potentially new target for therapeutic intervention in cervical cancer patients.

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