Abstract. Plants used in folklore medicine continue to be an important source of discovery and development of novel therapeutic agents. In the present study, we determined the effects of crude aqueous extracts of a panel of medicinal plants on the growth and invasion of cancer cells. Our results showed that extracts of L. tridentata (Creosote Bush) and J. communis L. (Juniper Berry) significantly decreased the growth of MCF-7/AZ breast cancer cells. The latter as well as A. californica (Yerba Mansa) inhibited invasion into the collagen type I gel layer. Furthermore, the phosphorylation levels of extracellular signal-regulated kinase 1 and 2 (ERK1/2) decreased when the cells were exposed to aqueous extracts of L. tridentata, J. communis L. and A. californica. This study provides original scientific data on the anticancer activity of selected aqueous medicinal plant extracts used in traditional medicine.

Introduction
Since ancient times, plants, herbs and spices have been important resources in traditional medicine (1). The use of plants in the treatment of a variety of diseases, including cancer, has played a significant role in nearly every culture on earth and is the basis of modern medicine (2). Natural products are considered powerful sources of novel drug discovery and development. Their dominant role in anti-cancer chemotherapeutics is evident with approximately 74% being either natural products or natural product-derived (3,4). Cancer is a major health problem in the United States and other developed countries. Breast cancer, the most commonly diagnosed cancer among women, is the leading cause of death, resulting from the metastatic spread of primary tumors (5). Therefore, there is an ongoing need for both the improvement of current therapeutic strategies and the search for novel agents.

In a previous study, we investigated the effects of two medicinal plants, L. porteri (Osha) and A. californica (Yerba Mansa), used by Native American tribes, on the growth of human breast cancer cells. We reported that both the aqueous and ethanol extracts of A. californica, exert an extracellular signal-regulated kinase (ERK)-mediated growth inhibitory effect on MCF-7/AZ cells, whereas L. porteri showed no such activity (6). In the present study, we explored their effects on the invasiveness of human breast cancer cells and subsequently investigated changes in the activity level of ERK1/2. The aim of this study was to evaluate the anticancer activity of crude aqueous extracts of medicinal plants used in traditional medicine in order to further identify active components or potential lead compounds.

Materials and methods
Plant materials and preparation of extracts. L. porteri, A. californica, L. tridentata, J. communis L., R. idaeus L., C. annuum L. and T. vulgaris were obtained from local herb stores or collected in New Mexico, USA. Cold aqueous extracts of each plant were prepared as follows: Seventy-five grams of the dried plants were soaked in water for 24 h at room temperature. The mixtures were then filtered to remove particulate matter and lyophilized. Table I shows the obtained
yields of the different plant extracts. The resulting powders were stored in a desiccator at 4°C.

Cell culture. MCF-7/AZ is a variant of the human mammary carcinoma cell family MCF-7 (7). The cells were maintained on a tissue culture plastic substrate (Nunc) in a mixture of Dulbecco's modified Eagle's medium (DMEM) and HAMF12 (50/50) (Invitrogen, Carlsbad, CA, USA) supplemented with 250 IU/ml penicillin, 100 μg/ml streptomycin (Invitrogen) and 10% fetal bovine serum (FBS) (Invitrogen), at 37˚C in a humidified atmosphere containing 10% CO₂.

Assay for cell viability. Cell viability was tested in accordance with Romijn et al (8). Briefly, mitochondrial dehydrogenase activities were measured by an MTT-reagent (Sigma, St. Louis, MO, USA). Cells were seeded in microtiter plates at an initial density of 1.5x10⁴ cells in 200 μl culture medium and treated with increasing concentrations of each plant extract. In each experiment, eight wells were used to determine the mean optical density (OD) referring to the cell viability.

Assay for cell growth. Cells were seeded in microtiter plates at an initial density of 1.5x10⁴ cells in 200 μl culture medium and treated with increasing concentrations of each plant extract. After an incubation period of 4 days the amount of cell protein in each well was estimated with the sulforhodamine B (SRB) assay (9) or the cells were counted with a hemacytometer (Hausser Scientific, Horsham, PA, USA).

Collagen type I invasion assay. This was performed as described previously (10). Briefly, six-well plates were filled with 1.25 ml neutralized type I collagen (0.09%) (Upstate Biotechnology, Lake Placid, NY, USA) and incubated for 1 h at 37˚C to allow gelification. Non-invasive MCF-7/AZ cells were pretreated with ET-18-Ome for 24 h in order to become invasive into collagen type I (11) and served as the control for invasiveness as compared to untreated MCF-7/AZ cells. Single cell suspensions were prepared with trypsin/EDTA, mixed with the different extracts, seeded on top of collagen type I gel and cultured at 37°C for 24 h. The number of cells penetrating into the gel or remaining at the surface was counted in 12 fields of 0.157 mm², using an inverted microscope controlled by a computer program. The invasion index expresses the percentage of invading cells over the total number of cells.

Western blotting. Cell lysates were made from 70% confluent cultures. The cells were treated for specified amounts of time with aqueous extracts of J. communis L. and T. vulgaris. After incubation, the cells were washed three times with PBS and lysed in 0.5 ml lysis buffer containing 1% Triton X-100, 1% NP-40 and the following protease inhibitors: Aprotinin (10 μg/ml), leupeptin (10 μg/ml), PMSF (1.72 mM), NaF (100 mM), NaVO₃ (500 μM) and Na₄P₂O₇ (500 μg/ml). Protein concentration was determined using the Pierce BCA protein assay kit (Pierce, Rockford, IL, USA). Aliquots of the lysates containing the same quantity of proteins were boiled for 5 min in sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) sample buffer containing 5% ß-mercaptoethanol, electrophoresed on 7.5% SDS-PAGE and transferred to PVDF membranes. After the transfer, the membranes were incubated with primary anti-phospho-ERK1/2 (1:1000) (Cell Signaling Technologies, Beverly, MA, USA), followed by incubation with a secondary biotinylated antibody (1:1000) and development by the enhanced chemiluminescence (ECL) detection kit (Vector Labs, Burlingame, CA, USA). The membranes were imaged and analyzed on the BioChemi System and analysis software (UVI, Upland, CA, USA).

Statistics. All treatments were matched and carried out at least 3 times. The Student's t-test (95%) was used for the statistical evaluation of the SRB, ERK1/2 phosphorylation and collagen type I invasion assay. The levels of phospho-ERK1/2 were quantified using Scion Image statistical software (Scion Corporation, Frederick, MD, USA).

Results

Effects of plant extracts on cell viability. The cytotoxic activity of each aqueous plant extract was determined on MCF-7/AZ breast cancer cells after 24-h exposure. A concentration-dependent decrease in cell viability was observed for the extracts of R. idaeus L. and J. communis L., whereas no significant changes were detected for C. annuum L., T. vulgaris and L. tridentata (data not shown). The cytotoxic effects of L. porteri and A. californica were previously determined (6).
For further experiments, the OD<sub>80</sub> values were used, leaving ~80% of the cells viable (Table II).

Effects of plant extracts on cell growth. SRB assay. Aqueous extracts of <i>J. communis</i> L. and <i>L. tridentata</i> markedly inhibited the growth of MCF-7/AZ cells in a concentration-dependent manner (Fig. 1A). All the other extracts showed no effect on the growth of MCF-7/AZ cells (data not shown).

Cell counting. MCF-7/AZ cells exposed to extracts of <i>J. communis</i> L. and <i>L. tridentata</i>, at 50 and 180 μg/ml (80% viability), respectively, showed a 30% decrease in cell proliferation as compared to the control cells (Fig. 1B), whereas no anti-proliferative effect was observed for the other extracts, as shown for <i>R. idaeus</i> L. and <i>T. vulgaris</i>. These findings are consistent with the results obtained from the SRB assay (Fig. 1A).

Effects of plant extracts on invasion into collagen type I gel layer. MCF-7/AZ cells are non-invasive into the collagen type I gel layer. ET-18-OMe treatment, however, induces invasion into collagen type I (11). <i>J. communis</i> L. and <i>A. californica</i> significantly reduced the induced invasion of MCF-7/AZ cells into collagen type I. No such effect was observed for the other plant extracts (Fig. 2).

Time-dependent effects of plant extracts on phosphorylation of ERK1/2. Exposure of MCF-7/AZ cells to aqueous extracts of <i>J. communis</i> L. and <i>L. tridentata</i> significantly decreased the phosphorylation levels of ERK as compared to the controls (Fig. 3). No changes in the phosphorylation levels were
Figure 2. Anti-invasive effects of aqueous extracts of *J. communis* L. (Juniper Berry) and *A. californica* (Yerba Mansa) ET-18-OHinduced invasive MCF-7/AZ cell suspensions were seeded on top of collagen type I gel, exposed to each plant extract in concentrations as determined by the MTT assay (Table II) and cultured at 37°C for 24 h. The invasion index expresses the percentage of invading cells into collagen type I over the total number of cells. The controls included untreated MCF-7/AZ and ET-18-OH-treated (invasive) MCF-7/AZ cells. The bars and flags indicate the mean values and standard deviations. The asterisks indicate statistical differences from the ET-18-OH-treated MCF-7/AZ cells.

Figure 3. Effects of aqueous extracts of *J. communis* L. (Juniper Berry) and *L. tridentata* (Creosote Bush) on ERK1/2 phosphorylation. MCF-7/AZ cells were exposed to 50 μg/ml and 180 μg/ml, respectively of *J. communis* L. and *L. tridentata* extracts for specified amounts of time. Whole cell lysates, containing 30 μg protein, were analyzed by 7.5% SDS-PAGE and immunoblotted with antibody against activated ERK (p-ERK, Thr202/Tyr204). The membranes were stripped at 50°C for 30 min in stripping buffer (100 mM β-mercaptoethanol, 2% SDS, 62.5 mM Tris-HCl pH 6.8) and rebotted with anti-ERK (data not shown). Scion image densitometry analysis of bands of activated ERK (p-ERK, Thr202/Tyr204), is expressed as the percentage of unexposed cells. The bars and flags indicate the mean values and standard deviations. The asterisks indicate statistical differences from the controls.
observed when the cells were exposed to the other aqueous extracts (data not shown). The total levels of ERK were unaltered upon exposure (data not shown).

Discussion

Historically, plants, herbs and spices were a folkloric source of medicinal agents, and as modern medicine expanded, many useful drugs were developed from lead compounds discovered from medicinal plants. This approach has provided leads against various pharmacological targets, including cancer, HIV, malaria and pain, and remains an important route to new pharmaceuticals (1-4,12).

In this study, we analyzed the effects of several medicinal plant extracts on the proliferation and invasion of MCF-7/AZ breast cancer cells. Aqueous extracts prepared from each medicinal plant showed no significant cytotoxic activity at concentrations lower than 50 μg/ml. MCF-7/AZ cells exposed to aqueous extracts from J. communis L. and L. tridentata potently inhibited the growth of MCF-7/AZ cells compared to unexposed cells as determined by SRB studies. The growth-inhibitory effect of L. tridentata has previously been reported (13) and is attributed to its major phenolic constituent, nordihydroguaiaretic acid (NDGA) (14). Certain anticancer properties of NDGA are related to the inhibition of receptor tyrosine kinases (RTKs), the insulin-like growth factor receptor (IGF-1R) and the c-erbB2/HER2/neu (HER2/neu) receptor (15). This correlates with our findings, namely with the decreased phosphorylation levels of ERK, since ERK is a downstream effector molecule of both receptors (16,17).

Additionally, this compound is a general inhibitor of lipoxygenases (18), the latter playing a crucial role in cancer cell survival and apoptosis, through the release of cytochrome c and the activation of specific caspases (19). Since extracts of J. communis L. inhibited the growth of cancer cells, we speculated that changes in ERK activity could be responsible for the growth inhibitory effects, given the fact that many plant-derived components modulate ERK activities in order to elicit their anti-neoplastic actions (20-22). A recent study also reveals that extracts of J. communis L. also inhibit 12-lipoxygenases (23). The possibility that J. communis L. and L. tridentata influence cell survival through the inhibition of lipoxygenases and the subsequent activation of caspases (19,24) was ruled out since no differences in caspase activities were observed (data not shown). Our findings revealed that J. communis L. and A. californica efficiently inhibited invasion, which could also be ascribed to the inhibition of the ERK activity (6). There is increasing evidence that ERK is an important mediator in the acquisition of a malignant phenotype, due to its role in multiple branching signaling pathways. Consequently, the specific blockade of ERK results not only in an anti-proliferative effect but also in an anti-invasive effect (27).

Although aqueous extracts of L. tridentata decreased ERK phosphorylation levels and earlier study has indicated that NDGA inhibits the expression of the intracellular adhesion molecule-1 (ICAM-1) on tumor cells (28) and is thus valuable for the treatment of metastasis, we did not find any inhibitory effect of L. tridentata on invasion. An explanation could be, that despite the presence of NDGA in aqueous extracts of L. tridentata, the activity of the extract can not be attributed to NDGA itself but rather to other components or influences of NDGA on the activity of other compounds present. A similar assumption was made for the extracts of L. divaricata and NDGA (29).

To the best of our knowledge, this is the first publication demonstrating that J. communis L. inhibits the growth of cancer cells, and that both J. communis L. and A. californica exert an anti-invasive effect. In addition, these results indicate that aqueous extracts of J. communis L. and A. californica comprise active components or potential leads that could be useful in cancer treatment. Studies along this line are currently being carried out in our laboratories.

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References

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