Norepinephrine-induced invasion by pancreatic cancer cells is inhibited by propranolol

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Abstract. Active migration and invasion by cancer cells are a prerequisite for the development of metastases. Recent studies have shown that neurotransmitters are involved in the regulation of cancer cell invasion via β-adrenoceptors (β-ARs). However, little is known regarding the effect of neurotransmitters on pancreatic cancer cells. The aim of our study was to examine the regulative effect of norepinephrine (NE), which belongs to the group of classical neurotransmitters, on the invasiveness of pancreatic cancer cells and the therapeutic effect of the β-blocker, propranolol, on them. The human pancreatic cancer cell lines, Miapaca-2 and Bxpc-3, were selected for this study, and in both cell lines, β1-AR and β2-AR expression was determined by RT-PCR and Western blotting. The invasiveness of pancreatic cancer cells was examined using the Matrigel invasion assay. The concentrations of MMP-2, MMP-9, and VEGF in the culture medium and in the cancer cells were examined by ELISA and RT-PCR, respectively. We observed that NE promoted the invasiveness of Miapaca-2 cells in a concentration-dependent manner, and NE increased the expression of MMP-2, MMP-9, and VEGF. However, these effects could be inhibited by the β-blocker, propranolol. In conclusion, the development of metastases is not only genetically determined, but is also influenced by NE, which is one of the signal substances present in the tumor environment. This study also provides experimental evidence for the use of β-blockers in the chemoprevention of pancreatic cancer metastasis.

Introduction

More than 90% of deaths from cancer do not result from the primary tumor, but from the development of metastases (1).

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Therefore, a vital aim in the treatment of cancer should be to inhibit the spread of tumor cells, including tumor cell migration and invasion, thus inhibiting the development of metastases. However, the complex process and mechanism of metastasis is not well understood to date, but that genetic alterations of the neoplastic cells as well as host factors in the tumor environment are believed to be involved. Genetic mutations theory has been developed by Fearon and Vogelstein for colorectal tumors (2). This theory is supported by the finding that mutations of the tumor suppressor gene, MADH4, occur more frequently in metastatic tumors (3). Host factors promoting cancer metastatic development is based on the finding that signal substances such as ligands to G protein-coupled receptors are capable of regulating the migratory and invasive activity of tumor cells, which is similar to the recruitment and homing of leukocytes in the immune system (4,5). These ligands predominantly consist of 2 groups, namely chemokines and neurotransmitters (6,7).

Norepinephrine (NE) belongs to the group of classical neurotransmitters (6), and it is one of the most potent known stimulators of tumor cell migration (8). Recently, the studies of Sood et al (9) and Lutgendorf et al (10,11) have shown that NE may influence the progression of ovarian cancer by modulating the expression of matrix metalloproteinases (MMPs) and the angiogenic cytokine, VEGF, in ovarian cancer cells. Studies in mice support these observations (12). It has also been indicated that NE can activate the migration of carcinoma cells from cancers of the breast (13), colon (14), and prostate (15). Furthermore, the NE-induced migration of these carcinoma cells has been shown to occur via β-adrenoceptors (β-ARs) (13-15).

More than 200,000 people die annually from pancreatic cancer in the world, and its 5-year survival rate is still below 5% (16). The incidence of pancreatic cancer is almost equal to its mortality rate. More than half of pancreatic cancer patients have distant metastasis. The morbidity and mortality from pancreatic cancer is conspicuously associated with metastasis (17). No substantial treatment improvements have been made, and the treatments have little effect on prolonging survival time. The expression of β1-AR and β2-AR in human pancreatic cancer cell lines and their role in the stimulation of cell proliferation were first described in 2001 (18), and these findings were later extended to show that β-ARs signaling in...
these cells included cAMP-dependent transactivation of the EGFR pathway (19). In our present study, we found that the human pancreatic cells, Miapaca-2, Bxpc-3 and PC-2, expressed both β₁-AR and β₂-AR (20). However, the role of β-ARs in the pancreatic cancer cells is not completely understood, and little is known regarding the effect of NE on pancreatic cancer cells. Therefore, we addressed whether NE is capable of stimulating metastatic invasion by pancreatic cancer cells and whether this effect can be inhibited by treatment with clinically established β-blockers.

We found that pancreatic cell lines in our present study expressed β-ARs. Previous studies have indicated that NE can promote the migration and invasion of carcinoma cells via β-ARs. Therefore, we hypothesized that NE, which belongs to the group of classical neurotransmitters, may play a role in the metastasis of pancreatic cancer, and β-blockers may be used for the chemoprevention of pancreatic cancer metastasis.

Materials and methods

Cell line and culture conditions. Human pancreatic cancer cell lines (Miapaca-2 and Bxpc-3) were stored in the Department of Hepatobiliary Surgery, the First Affiliated Hospital of Xi'an Jiaotong University. The cells were cultured in DMEM (Invitrogen, Carlsbad, CA, USA) supplemented with 10% FBS (Invitrogen), penicillin (100 U/ml) and streptomycin (0.1 mg/ml).

Matrigel invasion assay. An invasion assay was performed with a Millicell invasion chamber (Millipore, Billerica, MA, USA). The 8-μm pore inserts were coated with 15 μg of Matrigel (Becton-Dickinson Labware, Bedford, MA, USA). The cells were first seeded in 12-well plates at a concentration of 2.5x10⁵ per well and were cultured for 24 h with norepinephrine (Sigma, St. Louis, USA) at concentrations of 0.1, 1, and 10 μM. For the blocking experiments, 1 μM propranolol (Sigma) was added to the cell cultures 1 h before the addition of 10 μM norepinephrine. Normal culture medium was added at the bottom chamber to induce the cancer cell lines. Cells (5x10⁵) which were pretreated were seeded in the top chamber. The Matrigel invasion chamber was incubated for 24 h in a humidified tissue culture incubator. Non-invading cells were removed from the top of the Matrigel with a cotton-tipped swab. Invading cells on the bottom surface of filter were stained with Crystal Violet (Boster Biological Technology Ltd., Wuhan, China). Invasion ability was determined by counting the stained cells.

ELISA assays. The concentrations of total MMP-2, total MMP-9, and VEGF from the pancreatic cancer cell lines in the conditioned medium were detected using a commercial ELISA. The serum-free conditioned medium from the cultures were harvested and collected at 24 h following exposure to norepinephrine with or without propranolol. The supernatant was microfuged to remove the debris, and the remainder was then stored at -80°C. The samples were thawed only once for analysis. Secretion of MMP-2, MMP-9 and VEGF (Peprotech, Rocky Hill, NJ, USA) were later quantified with ELISA kits per the manufacturer's instructions. Samples were assayed in duplicate.

Reverse transcription-PCR. Total RNA was isolated from the Miapaca-2 and Bxpc-3 cells using TRIzol reagent (Invitrogen Life Technologies) according to the manufacturer's instructions. DNase was used to remove the contaminating genomic DNA after RNA purification. cDNA synthesis was performed by heating 3 μg of total RNA with 0.5 μg of random primers and dNTP (final concentration, 0.5 mM) at 65°C for 5 min. Reverse transcription was then performed at 42°C for 1 h with this mixture at a final volume of 20 μl containing 0.5 M Tris-HCl (pH 8.0), 0.5 M potassium chloride, 0.05 M MgCl₂, 40 U RNase inhibitor (MBI Fermentas Life Sciences, Foster, CA), 200 U M-MLV reverse transcriptase (Gibco), and 10 mM dithiothreitol in RNase-free water. PCR amplification was performed on 2 μl of the synthesized cDNA under the following conditions: 94°C for 3 min, 30 cycles of denaturation for 45 sec at 94°C, 45 sec of annealing at 58°C, elongation at 72°C for 35 sec, and extension at 72°C for 5 min. Mixtures containing 9 μl of the resulting PCR fragments and 1 μl of 10X loading buffer were electrophoresed on a 1.5% agarose gel in 1X TAE buffer at 110 V for 30 min at room temperature. The resulting bands on the gel were photographed and analyzed using a gel-imaging analyzer. Levels of gene expression were presented as the ratios of densities between PCR products and β-actin in the same sample. All experiments were repeated three times.

The primers were as follows: MMP-2: forward, 5'-CCG TCC CCC ATC ATC AAG TTC-3' and reverse, 5'-GCA GCC TGT CGG TGA GAT TGG-3' (90 bp). MMP-9: forward, 5'-CTG GCC TGT CCT GCC TCC-3' and reverse, 5'-GCT GCC TGT CGG TGA GAT TGG-3' (111 bp). VEGF: forward, 5'-CTG GCC TGT CCT GCC TCC TTC G-3' and reverse, 5'-CCT GCC TTC TCT TCC TCT TCT TCC-3' (140 bp). β₁-AR: forward, 5'-GGG AGA AGC ATT AGG AGG G-3' and reverse, 5'-CAA GGA AAG CAA GGT GGG-3' (270 bp). β₂-AR: forward, 5'-CAG CAA AGG CAA GTG GGG-3' (334 bp). β-actin (used as a loading control): forward, 5'-ATC GTG CTC GAG ATT AAG GAG AAG-3' and reverse, 5'-AAG TAA TGG CAA AGT AGC G-3' (334 bp).

Protein extraction and Western blotting. Total protein was isolated from 1x10⁶ cells with 200 μl of ice-cold lysis buffer containing 1% Nonidet P-40 (NP-40), 50 mM Tris (pH 7.4), 150 mM NaCl, 0.1% sodium dodecyl sulfate (SDS), 0.5% deoxycholate, 200 mg/ml phenylmethylsulfonyl fluoride (PMSF), and 50 mg/ml aprotinin. Insoluble materials were removed by centrifugation at 15,000 x g for 15 min at 4°C. The concentration of the extracted protein was measured spectrophotometrically with Coomassie G-250. Cell lysates containing equal amounts of protein were electrophoretically resolved on a denaturing SDS-polyacrylamide gel (12%), and electrotransferred onto nitrocellulose membranes. The membranes were initially blocked with 5% non-fat dry milk in Tris-buffered saline (TBS) for 2 h and then incubated in β₁-adrenergic receptor (1:500) or β₂-adrenergic receptor (1:500)
primary antibodies (Abcam, Cambridge, MA) at 4˚C over-night and secondary antibodies for 2 h at room temperature. The protein bands specific for antibody were visualized by enhanced chemiluminescence associated fluorography. The Western blotting was graded positive if the band of interest was present at the expected molecular weight corresponding to each marker protein. All analyses were done in duplicate.

Statistical analysis. All statistical analyses were performed using the SPSS13.0 software. The results are presented as mean ± SD of three replicate assays. Differences between the groups were assessed by analysis of variance (ANOVA). P<0.05 was considered to indicate statistical significance.

Results

Expression of β-ARs in the human pancreatic cancer cells. RT-PCR analysis revealed that both β₁-AR and β₂-AR were expressed by both Miapaca-2 and Bxpc-3 cell lines (Fig. 1A). Western blot analysis showed β₁-AR immunoreactivity was visualized as a single band that migrates at ~64 kDa, whereas β₂-AR migrates at 95 kDa and the β₂-AR protein yielding the more prominent band (Fig. 1B).

Increased cell invasion of human pancreatic cancer cells by NE is inhibited by β-blocker, propranolol. The effect of treatment with NE on pancreatic cancer cell line Miapaca-2 was studied in vitro. The number of invasive cells pretreated with NE increased in a concentration-dependent manner (Fig. 2A-D). Significant effect on cell invasiveness with a ~2.1-fold and ~2.8-fold increase as compared to control in case of 1 μM and 10 μM of NE, respectively. To further determine whether the β-ARs mediated the effect of NE on the invasiveness of the pancreatic cancer cells, we pretreated the Miapaca-2 cells with the broad-spectrum β-blocker, propranolol (1 μM), 1 h before the addition of 10 μM NE for 24 h. Our results showed that propranolol reduced the number of invaded pancreatic cancer cells of NE-stimulated by ~3.5-fold (Fig. 2). These results suggest that exogenous NE stimulated invasiveness by pancreatic cancer cells and propranolol blocked NE-mediated stimulation of the invasiveness of human pancreatic cancer cells.
Propranolol inhibits NE-mediated increase of MMP-2, MMP-9 and VEGF levels in human pancreatic cancer cells.

We examined the effect of NE on the secretion of MMP-2, MMP-9, and VEGF in Miapaca-2 cells. Our results showed that NE dramatically increased the levels of secreted MMP-2, MMP-9 and VEGF at 24 h post treatment (Fig. 3). NE (10 μM) induced a ~3.6-fold, ~4.1-fold and ~2.9-fold increase in the MMP-2, MMP-9, and VEGF levels in the Miapaca-2 supernatant at 24 h post-stimulation. The RT-PCR results showed that the MMP-2, MMP-9, and VEGF gene expression were significant up-regulation after treatment with 10 μM NE for 24 h (Fig. 4). The data suggest that the increase in the levels of MMP-2, MMP-9, and VEGF protein was a result of transcriptional changes. Since the β-blocker, propranolol, has been shown to have inhibitory effects on NE-mediated increases in cell invasiveness of human pancreatic cancer cells, we also explored its ability to inhibit MMP-2, MMP-9, and VEGF. Interestingly, our results showed that propranolol reduced the increased levels of secreted MMP-2, MMP-9 and VEGF levels in NE-stimulated Miapaca-2 cells (Figs. 3 and 4). This suggests that propranolol abrogated NE-induced up-regulation of MMP-2, MMP-9, and VEGF release, indicating that the action of NE on pancreatic cancer cell invasiveness was through β-ARs.

Discussion

In this study, we addressed the effect of NE, which belongs to the group of classical neurotransmitters, on the invasiveness of pancreatic cancer cells, which is a critical component of the metastatic cascade. Our data show that NE can significantly enhance the capacity of the pancreatic cancer cells to invade the extracellular matrix that is characteristic of the basement membrane. Furthermore, we showed that NE can up-regulate the production of MMP-2, MMP-9, and VEGF in the Miapaca-2 cells. These factors, which are responsible for the invasiveness and angiogenic responses, facilitate penetration of the extracellular matrix of the pancreatic cancer cells. We also showed that treatment with the β-blocker, propranolol, to block the binding of NE to β-ARs suppresses the invasiveness of Miapaca-2 cells and abrogates the up-regulation of MMP-2, MMP-9, and VEGF. These data suggest that NE plays a role in the invasiveness of pancreatic cancer cells, and β-blocker may be used for the chemoprevention of pancreatic cancer metastasis.

Tumor cell migration and invasion, the prerequisite for metastasis development, are not only merely genetically determined but are also distinctly regulated by signal substances in the environment, including chemokines and neurotransmitters (4-7). It is well known that chemokines regulate the migratory activity and homing of leukocytes (21). Recently, it has become evident that chemokines are also involved in the development of metastasis and localization of cancer cells (22,23). Neurotransmitters are signal substances that traditionally have functions in both the central and peripheral nervous systems. The pancreas is richly supplied with nerves. Deriving from different locations, nerve fibers of different types intermingle as they enter or leave the pancreas, frequently closely adherent to pancreatic arteries (24). Current studies have revealed that neurotransmitters have direct influence on the migratory activity and invasiveness of tumor cells (6). NE belongs to the group of classical neurotransmitters. It can modulate carcinogenesis induced by a tobacco-specific nitrosamine in the lungs of hamsters (25). It can also activate the migration and invasion of carcinoma cells in cancers of the breast (13) and the colon (14). Furthermore, NE induces chemotaxis in breast carcinoma cells (13), and metastases of small-cell lung carcinoma (SCLC) are very common in the NE-producing adrenal glands and the brain (26). Overexpression of NE has recently been described in the majority of investigated human pancreatic cancers (27), a finding that strengthens the significance of NE on cancer progression. In our study, we observed that exogenous NE stimulates the
invasiveness of the pancreatic cancer cells in a concentration-dependent manner. This indicated that the invasiveness of pancreatic cancer cells was distinctly regulated by NE. NE is a neurotransmitter that is also released in stress reactions. The long-lasting elevation of catecholamines attributable to chronic stress is known to be a risk factor for cancers (28). Thus, we also provide evidence for a functional link between chronic stress and its influence on pancreatic cancer cells.

The process of tumor cell penetration of the host basement membrane consists of attachment, matrix dissolution, motility, and penetration (29). The ability of tumor cells to invade the extracellular matrix plays an important role in invasion and metastasis. MMPs are key factors in the degradation of the components of the extracellular matrix, such as collagen, laminins, fibronectins, elastins, and the protein core of proteoglycans (30). The coexpression of MMPs and VEGF has been described in many human cancers. High levels of VEGF and MMPs have been described in patients with ovarian cancer (31). The level of VEGF has been shown to be positively correlated with those of MT1-MMP, MMP-2, and MMP-9 in human glioblastomas (32). Studies have indicated that levels of MMP-9 and VEGF are positively correlated in head and neck squamous cell carcinoma (33). In our study, we found that NE had a role in modulating the expression of MMP-2, MMP-9, and VEGF in pancreatic cancer cells. The NE-mediated up-regulation of MMP-2, MMP-9, and VEGF contributed to the aggressiveness of the highly metastatic forms of pancreatic cancer cells.

$\beta_1$-AR and $\beta_2$-AR were expressed in the pancreatic cancer cells. Evidence supporting the role of these receptors in the NE-dependent effect is provided by our results showing that propranolol inhibited the NE-dependent invasiveness of cells and the up-regulation of MMP-2, MMP-9, and VEGF release. B-ARs antagonists have been used as therapies for hypertension and the prevention of heart attacks for many years. Increasing evidence suggests that patients with pancreatic cancer share many risk factors with cardiovascular disease patients, such as smoking and a high-fat diet (34). Therefore, our findings might open new pharmacological possibilities for the preventive treatment of pancreatic cancer, i.e., to delay or to inhibit the progression of the disease with regard to invasion and the development of metastases. However, further approaches to inhibit pancreatic cancer cells invasion and the relevant signal transduction pathways remain to be elucidated, and whether NE leads to organ-specific metastasis to other NE-rich organs remains to be investigated.

In summary, our results show that NE, which belongs to the group of classical neurotransmitters, can directly enhance the invasive potential of pancreatic cancer cells via B-ARs by mediating the up-regulation of MMP-2, MMP-9, and VEGF, and this effect can be inhibited by the B-blocker, propranolol. These results support the theory that the development of metastases is not only genetically determined but is also influenced by signal substances in the tumor environment. This study also provides experimental evidence for the use of B-blockers for chemoprevention of pancreatic cancer metastasis.

References