Breast MALT lymphomas: A clinicopathological and cytogenetic study of 9 cases

GIUSEPPINA LIGUORI1, MONICA CANTILE1, MARGHERITA CERRONE1, ELVIRA LA MANTIA1, MAURIZIO DI BONITO1, FABRIZIO ZANCONATI2, MARIA PIA CURCIO1, GABRIELLA AQUINO1, ANNA LA MURA3, ROSA GIANNATIEMPO3, ANNA ROSARIO DE CHIARA4, ANGELA LOMBARDI5, GERARDO BOTTI1, ANTONIO D’ANTONIO6, MICHELE CARAGLIA5 and RENATO FRANCO1

1Pathology Unit, National Cancer Institute, Pascale Foundation, Naples; 2UCO Clinical Surgery, University of Trieste, Trieste; 3Pathology Unit, Scafati Hospital, Salerno; 4Pathology Unit, Villa Betania Hospital; 5Department of Biochemistry and Biophysics, Second University of Naples, Naples; 6Department of Pathological Anatomy and Oncology, A.O.U. ‘San Giovanni di Dio e Ruggi d’Aragona’, Salerno, Italy

Received March 13, 2012; Accepted May 8, 2012

DOI: 10.3892/or.2012.1942

Abstract. Primary breast mucosa-associated lymphoid tissue (MALT) lymphomas are uncommon and restricted diagnostic criteria should be used to exclude breast involvement by systemic lymphomas. The molecular pathogenesis of primary breast MALT lymphomas is not clear because of the rarity of the disease. Generally the molecular studies of MALT lymphoma in extranodal sites have shown the presence of different chromosomal aberrations, mutually exclusive with substantial differences in their frequency relatively to topographic localization. Few cases of breast MALT lymphomas in the literature have been assessed for MALT lymphoma-associated translocations and BCL10 expression, underlying their rarity in primary breast MALT lymphomas. In our study, we analyzed a series of nine cases of primary breast MALT lymphomas. FISH results showed evidence of MALT1 gene rearrangements in four primary breast lymphomas, in particular three cases with t(11;18)(q21q21) and one case with t(14;18)(q32;q21). In addition, BCL10 gene rearrangement was not observed. There was no evidence of BCL10 gene translocation in any of the neoplasms assessed. Our data indicate that MALT1 gene rearrangements are not rare in primary breast MALT lymphoma in contrast with results of previous series. Finally, t(11;18) has been observed to be significantly associated with high intensity cytoplasmic BCL10 expression underlying cross-talk between MALT1 and BCL10 pathways in the pathogenesis of MALT lymphomas.

Introduction

Malignant lymphomas observed in the breast are most commonly non-Hodgkin lymphomas. They could be primary breast lymphoma (PBL) or most frequently a sign of systemic disease (1,2). The PBL represent 0.38-0.70% of all non-Hodgkin lymphomas (NHL), 1.7-2.2% of all extra nodal NHL and only 0.04-0.5% of all breast cancer (1-3). The WHO criteria for the diagnosis of PBL are clinicopathological demonstration of lymphomatous infiltration within breast tissue with or without involvement of axillary lymph nodes, but in absence of systemic disease (1,4). The most common histotypes of PBL are diffuse large B-cell lymphoma and extra nodal marginal zone B-cell lymphoma of mucosa-associated lymphoid tissue MALT-type (MALT lymphomas) (1,2).

The molecular studies of MALT lymphoma in extra nodal sites have shown the presence of different chromosomal translocations that appear to be mutually exclusive with substantial differences in their frequency in relation to extra nodal anatomic sites. These translocations include the t(11;18)(q21q21), t(14;18)(q32;q21), t(1;14)(q22;q32), and the most recently described, t(3;14)(p14.1;q32) (5-7). These translocations result in the production of a chimeric protein (API2-MALT1) or in BCL10, MALT1, FOXP1 genes over expression, because of direct control of IgH promoter (8-10).

The purpose of this study is the evaluation of chromosomal rearrangements t(11;18)(q21q21), t(14;18)(q32;q21), t(1;14)(q22q32) and BCL10 expression in a series of nine cases of primary breast MALT lymphomas.

Materials and methods

Clinical information. Cases were selected from the pathological files of the Cardarelli Hospital and National Tumor Institute Pascale, Naples, and University of Trieste, from January 1993 to December 2010. The WHO criteria to establish the diagnosis of PBL have been strictly applied. Thus cases included in our study are characterized by the following
features: i) histologically the lymphomatous infiltrate should demonstrate a close relationship with the breast parenchyma with or without ipsilateral axillary lymph node involvement; ii) no evidence of systemic disease after staging; iii) imaging studies clearly identified the neoplasm to be within the breast in absence of other localization. Clinical information was recovered from clinical files. A total of 9 cases of MALT-like lymphomas were identified.

Specimens had been routinely fixed and processed. Haematoxylin and eosin (H&E) stained sections and appropriate immunohistochemical staining were performed in order to confirm diagnosis of MALT-like PBL (Table I). As part of this study, additional immunostaining has been performed using antibodies specific for BCL10.

Stained sections were evaluated by four different pathologists (R.F., A.D., M.D.B., G.B.) using uniform criteria. Discrepancies were resolved through simultaneous evaluation and discussion of the results. Single-marker expression was recorded as negative/positive and high/low level, after consideration of the expression in reactive compared with tumoral cells and the specific cut-off of each marker. As proposed, cytoplasmic BCL10 expression was scored as strong when it was similar to tonsil centroblast positivity, moderate when similar to centrocyte positivity, and weak/absent when similar to tonsil mantle-zone positivity. Nuclear positivity has been also recorded (11).

Fluorescent in situ hybridization study. Tissue array sections from paraffin-embedded tissue were heated for 4 h at 62°C and immediately deparaffinized in two rinses of 100% xylene for 10 min each. The slides were then treated with 0.3 M sodium chloride and 0.03 M sodium citrate for 20 min at 80°C, and with 0.05 mg/ml proteinase for 10 min at 37°C. For t(11;18) (q21;q21) detection, we used LSI API2/MALT1 t(11;18) (q21;q21) dual-colour, dual-fusion translocation probe, and for detection of BCL10 translocation BCL10 FISH DNA Probe, Split Signal, and Histology FISH kit (Dako) were used. The cut-off value for the diagnosis of rearrangement involving IGH and MALT1 was 5.3%, which is above the mean percentage of cells with a false positive signal plus 3 SD, as assessed in tissue from reactive tonsils present in TMA. Moreover, IGH dual-colour break-apart rearrangement probes (Vysis Inc., Downers Grove, IL, USA) were applied to cells of all t(14;18)(q32;q21)-positive lymphomas to confirm the translocation.

The Spectrum Green-labelled LSI IGVH probe covers the entire IGH variable region, while the Spectrum Orange-labelled probe lies completely within the IGH locus. As a result of this probe design, any translocation with a breakpoint at the J segments, or within switch sequences, should produce separate orange and green signals. Additionally, FISH with centromere-specific probes for chromosome 18 (Vysis) was performed in all cases. The appropriate probe mix (10 ml) was applied to the tissue sections and covered with a coverslip. Both probe and target DNA were simultaneously denatured at 75°C for 5 min and incubated overnight at 37°C using the Hybrite System. Post-hybridization washes were performed according to the `rapid wash protocol’ provided by Vysis. Slides were counterstained with 406-diamidino-2-phenylindole 2HCl (DAPI). FISH was performed according to the manufacturer's instructions (Vysis). FISH data were collected using an Olympus BX 61 fluorescence microscope equipped with a cooled black-and-white camera controlled by the associated software (Olympus, Italy).

Statistical analysis. The Pearson's χ² test was used where appropriate, to establish whether there were any relationships between the frequencies of different markers included in this study. Differences were considered to be significant for values of P<0.05. All statistical analyses were performed using the SPSS 98 v.12 program.

Table I. Immunohistochemical antibodies.

<table>
<thead>
<tr>
<th>Antigen clone</th>
<th>Source</th>
<th>Dilution</th>
<th>Reactivity</th>
<th>Threshold</th>
<th>Internal control</th>
</tr>
</thead>
<tbody>
<tr>
<td>CD20 (clone L-26)</td>
<td>Dako</td>
<td>1:100</td>
<td>Positive/negative</td>
<td>Any positive neoplastic cells</td>
<td>Reactive lymphocyte</td>
</tr>
<tr>
<td>CD10 (clone56C6)</td>
<td>Novocastra</td>
<td>1:10</td>
<td>Positive/negative</td>
<td>Any tumoral cell positive</td>
<td>GC* B cells</td>
</tr>
<tr>
<td>Bcl6 (clonePG-B6p)</td>
<td>Dako</td>
<td>1:10</td>
<td>Positive/negative</td>
<td>&gt;10% neoplastic cells</td>
<td>GC* B cells</td>
</tr>
<tr>
<td>CK (clone AE1/AE3)</td>
<td>Dako</td>
<td>1:50</td>
<td>Positive/negative</td>
<td>Any positive neoplastic cells</td>
<td>Breast epithelial cell</td>
</tr>
<tr>
<td>Bcl10</td>
<td>Dako</td>
<td>1:200</td>
<td>Positive/negative</td>
<td>&gt;10% neoplastic cells</td>
<td>Reactive lymphocyte</td>
</tr>
<tr>
<td>CD3</td>
<td>Dako</td>
<td>1:25</td>
<td>Positive/negative</td>
<td>Any tumoral cell</td>
<td>Reactive lymphocyte</td>
</tr>
<tr>
<td>CD5</td>
<td>Novocastra</td>
<td>1:50</td>
<td>Positive/negative</td>
<td>&gt;10% positive cells</td>
<td>Reactive lymphocyte</td>
</tr>
</tbody>
</table>
Results

Clinical features. The main clinicopathological data are reported in the Table II. All patients were female, with mean age of 72 years (range: 39-93). In 6 cases right breast was involved. Mean time of follow-up was 51 months (range: 33-65) and at the end of follow-up, 2 patients were alive, 6 alive with disease and 1 dead from disease.

Histological findings. All nine cases of MALT lymphoma had a diffuse or vaguely nodular growth pattern. We have identified three types of lymphoid cells: i) small to medium lymphoma cells with irregular nuclear contours, inconspicuous nucleoli and pale-staining cytoplasm (centrocyte-like cells), prevalent in all cases; ii) small lymphoid cells with round nuclei, clumped chromatin and sparse cytoplasm, some with prominent plasmacytic differentiation admixed with mature plasma cells; iii) lymphoid cells with abundant pale or clear cytoplasm resembling monocytoid cells. These elements are present in a variable number in all cases. Lymphoepithelial lesions were present but were not prominent. Transformed lymphocytes resembling centroblasts or immunoblasts were present in all cases, but rarely formed small clusters or confluent sheets. Lymphoid follicle with reactive germinal centre was also present in breast tissue in two cases. Mitotic activity was low and necrosis areas were not observed (Fig. 1).

Figure 1. (a) Epithelial islands in a diffuse pattern of lymphoma proliferation, x10, hematoxylin and eosin stain; (b) Lymphoma proliferation surrounds epithelial ducts, x40; (c) Monocytoid differentiation of MALT lymphoma, x40; (d) Plasmacytoid differentiation in MALT lymphoma, x40; (e) CK immunostaining, x40, in trapped duct cells by lymphoma; (f) CD20 immunostaining, x40, diffuse neoplastic cells positivity; (g) BCL6 immunostaining, x40, BCL6 negativity in MALT lymphoma; (h) Ki67 immunostaining, x40, low proliferation index of lymphoma.
Immunohistochemical findings. Immunohistochemical results are summarized in Table II. Lymphoma cells express pan B-lymphocyte antigens CD20 and CD79a in all cases and show κ light chain restriction. CD43 was expressed in 8/9 cases, while CD5, CD10, BCL6 and CD10 that were usually negative. BCL10 was positive with cytoplasmic high intensity in 4 cases and negative in 5. No nuclear positive case was observed (Fig. 2).

Table II. Main clinicopathological data.

<table>
<thead>
<tr>
<th>Case no.</th>
<th>Gender</th>
<th>Age</th>
<th>Site</th>
<th>BCL10 IHC</th>
<th>t(11;18)</th>
<th>t(14;18)</th>
<th>t(1;14)</th>
<th>Status</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>F</td>
<td>71</td>
<td>R</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>AWD</td>
</tr>
<tr>
<td>2</td>
<td>F</td>
<td>82</td>
<td>R</td>
<td>1+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>AWD</td>
</tr>
<tr>
<td>3</td>
<td>F</td>
<td>93</td>
<td>R</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>AWD</td>
</tr>
<tr>
<td>4</td>
<td>F</td>
<td>73</td>
<td>R</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Alive</td>
</tr>
<tr>
<td>5</td>
<td>F</td>
<td>74</td>
<td>L</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>Dead</td>
</tr>
<tr>
<td>6</td>
<td>F</td>
<td>71</td>
<td>L</td>
<td>2+</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>AWD</td>
</tr>
<tr>
<td>7</td>
<td>F</td>
<td>72</td>
<td>R</td>
<td>2+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>AWD</td>
</tr>
<tr>
<td>8</td>
<td>F</td>
<td>73</td>
<td>L</td>
<td>2+</td>
<td>+</td>
<td>-</td>
<td>-</td>
<td>AWD</td>
</tr>
<tr>
<td>9</td>
<td>F</td>
<td>39</td>
<td>R</td>
<td>-</td>
<td>-</td>
<td>+</td>
<td>-</td>
<td>Alive</td>
</tr>
</tbody>
</table>

F, female; R, right breast; L, left breast; AWD, alive with disease.

Figure 2. (a) Negative BCL10 immunostaining, x40; (c) Moderate BCL10 immunostaining, x40; (e) Intense BCL10 immunostaining, x40; (b, d and f) FISH t(1;14), x100, integrity of BCL10 genes signals in all cases.
**FISH analysis.** t(11;18)(q21;q21) was present in 3 cases and t(14;18)(q32;q21) in one case. There was no evidence of (1;14) (q22;q32) in any of the lymphoma assessed (Fig. 2).

**Statistical analysis.** Only association between t(11;18) and high intensity cytoplasmic expression of BCL10 (P=0.046) was observed.

**Discussion**

MALT type extra nodal marginal zone B-cell lymphoma (MALT lymphoma) represents <10% of all B-cell lymphomas (1). They are most frequent in the gastrointestinal tract, salivary glands, ocular adnexa, lungs, thyroid and breast (8). Due to the lack of native lymphoid tissue in these organs, a chronic inflammatory stimulus or an autoimmune disease can generate mucosa-associated lymphoid tissue from which a MALT lymphoma could arise (1,2,3,5-9). Histologically MALT lymphomas show a case-to-case variability in the neoplastic cytotype. Usually we have observed small cell lymphomas constituted by centrocyte-like cells with irregular nuclei, monocytoid B-cell with pale to clear cytoplasm with round nuclei and more rarely plasmacytoid cells. The different histological features of the tumor cells do not impact on clinical prognosis (9). Generally MALToma have an indolent behavior and recurrence can occur after many years also in other extra nodal sites. Moreover, in some cases a progression to diffuse large B-cell lymphoma may occur (1,9). In particular in a recent work primary breast MALT lymphomas seem to have a more indolent behavior in respect to other PBLs (12).

The molecular events underlying the origin of a mucosa-associated lymphoid tissue (MALT) lymphoma and their progression and prognosis are largely unknown. However, some chromosomal translocations have been identified in a subset of MALT lymphomas, including the t(11;18)(q21;q21), t(14;18)(q32;q21), t(1;14)(p22;q32), and t(3;14)(p14.1;q32). These translocations are responsible for deregulation of MALT1, BCL10 and FOXP1 genes that cooperate in the NFκB anti-apoptotic pathway (13). Particularly, t(1;18)(q21;q21) and t(14;18)(q32;q21) generate chimeric transcripts of MALT1 gene respectively with the API2 and IgH gene, whereas t(1;14) (p22;q32) causes aberrant BCL10 expression with nuclear localization (5-9).

In addition, nuclear BCL10 expression also occurs in MALT lymphomas without t(1;14)(p22;q32), suggesting an important role for BCL10 in lymphoma development (14-17). All these abnormalities also appear to be mutually exclusive and their frequency correlates with the anatomic site. The translocation (11;18) is frequently described in gastrointestinal and pulmonary sites in contrast with presence of (14;18) in ocular adnexa, skin and salivary gland (1-7,10,11). Different series regarding the most common extra nodal sites of MALT lymphomas have been screened for the presence of the common translocations involving MALT1 gene and BCL10 expression with relation to progression of disease or prognosis (11,14-18). t(11;18)(q21;q21) is associated with advanced gastric MALT-lymphoma that expresses nuclear BCL10. Moreover, nuclear expression of BCL10 or nuclear factor κB predicts *Helicobacter pylori*-independent status of early-stage, high-grade gastric mucosa-associated lymphoid tissue lymphomas (17). Moreover, nuclear BCL10 expression characterizes a group of ocular adnexa MALT lymphomas with shorter failure-free survival (11).

While the frequency of these translocations has been studied in the most common extra nodal sites, their presence has not been well characterized in the breast presumably because these neoplasms are very infrequent. In a previous study, Talwakar et al (18) studied eight cases of primary MALT breast lymphoma and 14 cases of primary breast diffuse large B-cell lymphoma for MALT1 gene rearrangements through FISH and for BCL10, NF-κB p65, p50 using immunohistochemical methods. None of the cases showed MALT1 gene rearrangements and NF-κB activation was not demonstrated. Similar results were obtained by Streubel et al (7) in five additional cases of breast MALT lymphoma assessed for the t(11;18) and t(14;18) and by Mulligan et al in a small series of PBL MALT lymphomas assessed for MALT1 gene rearrangement (19). All these authors concluded that MALT1 gene rearrangements are absent or rare in primary breast MALT lymphoma compared to other extra nodal MALToma. In our series both the t(11;18) and t(14;18) were observed. Our results showed evidence of MALT1 gene rearrangements in four PBL: t(11;18)(q21; q21) was observed in 3 cases and t(14;18)(q32; q21) in one case. The expression of BCL10 was only cytoplasmic in 4 cases, with low/moderate intensity, while nuclear expression was not observed. No association with chromosomal aberration was observed.

In conclusion, our data indicated that MALT1 gene rearrangements are not rare in primary breast MALT lymphoma, in contrast with results of previous series, involving mainly t(11;18)(q21;q21).

**References**


