Enhancement of antitumor immunity by combining anti-cytotoxic T lymphocyte antigen-4 antibodies and cryotreated tumor lysate-pulsed dendritic cells in murine osteosarcoma

MASANORI KAWANO, ICHIRO ITONAGA, TATSUYA IWASAKI and HIROSHI TSUMURA

Department of Orthopaedic Surgery, Faculty of Medicine, Oita University, Oita 879-5593, Japan

Received August 16, 2012; Accepted October 30, 2012

DOI: 10.3892/or.2013.2224

Abstract. Immunotherapy with tumor lysate-loaded dendritic cells (DCs) is one of the most promising strategies to induce antitumor immune responses. However, the antitumor activity of cytotoxic T cells may be restrained by their expression of the inhibitory T-cell coreceptor cytotoxic T lymphocyte antigen-4 (CTLA-4). By relieving this restraint, CTLA-4-blocking antibodies promote tumor rejection, but the full scope of their most suitable applications has yet to be fully determined. In the present study, we offer proof of a preclinical concept in a C3H mouse osteosarcoma (LM8) model that CTLA-4 blockade cooperates with cryotreated tumor lysate-pulsed DCs in a primary tumor to prevent the outgrowth of lung metastasis. To evaluate immune response activation, we established the following four groups of C3H mice (60 mice in total): i) control immunoglobulin G (IgG)-treated mice; ii) tumor lysate-pulsed DC-treated mice; iii) anti-CTLA-4 antibody-treated mice and iv) tumor lysate-pulsed DC- and anti-CTLA-4 antibody-treated mice. The mice that received the tumor lysate-pulsed DCs and anti-CTLA-4 antibody displayed reduced numbers of regulatory T lymphocytes and increased numbers of CD8 T lymphocytes inside the metastatic tumor, inhibition of metastatic growth, a prolonged lifetime, reduced numbers of regulatory T lymphocytes in the spleen and high serum interferon-γ levels. Combining an anti-CTLA-4 antibody with tumor lysate-pulsed DCs enhanced the systemic immune response. In the present study, we investigated how immunotherapies that target the inhibitory pathways of Tregs using anti-CTLA-4 mAbs potentially synergize the effects of cryotreated tumor lysate-pulsed DCs to generate systemic antitumor immunity. We verify that, in contrast to tumor lysate-pulsed DC or CTLA-4 treatment alone, the combination therapy enhanced antitumor immunity.
immunity and slowed the growth of a secondary tumor from a large primary tumor by using a mouse metastatic osteosarcoma model.

**Materials and methods**

**Cell line.** LM8 cells, derived from Dunn osteosarcoma, were provided by the Riken BioResource Center (Saitama, Japan). The cells were maintained in complete medium consisting of RPMI-1640 supplemented with 10% heat-inactivated fetal bovine serum, 100 µg/ml streptomycin and 100 U/ml penicillin. Cells were cultured at 37˚C in 5% CO₂.

**Animals.** A total of 1x10⁶ LM8 cells (a murine osteosarcoma cell line) was hypodermically implanted into the subcutaneous gluteal region of 60 female C3H mice 6-8 weeks old. We purchased the C3H mice from Sankyo Labo, Inc. (Toyama, Japan) and housed them in a specific pathogen-free animal facility in our laboratory.

**Study design.** All animals developed tumors. The following 4 groups were established (Fig. 1): i) control immunoglobulin G (IgG) (control; n=15); ii) DCs exposed to cryotreated tumor lysates were injected twice a week into the subcutaneous contralateral gluteal region [DC(Ly); n=15]; iii) intraperitoneal injection of anti-CTLA-4 Ab was performed twice/week (anti-CTLA-4 Ab; n=15) and iv) DCs exposed to cryotreated tumor lysates were injected twice a week into the subcutaneous contralateral gluteal region and intraperitoneal injection of agonist anti-CTLA-4 antibody was performed twice/week [DC(Ly) + anti-CTLA-4 Ab; n=15]. This study was performed at the Department of Orthopaedic Surgery, Faculty of Medicine, Oita University, Oita, Japan. All experiments were performed under the guidelines for animal experiments as stipulated by the Oita University Graduate School of Medical Science.

**Generation of DCs.** Bone marrow-derived DCs were generated as described by Lutz and Rössner (22) with minor modifications. Briefly, erythrocyte-depleted mouse bone marrow cells obtained from flushed marrow cavities (1x10⁶ cells/ml) were cultured in complete medium with 20 ng/ml recombinant mouse granulocyte-macrophage colony-stimulating factor (GM-CSF) (PeproTech EC Ltd., London, UK) in 10-cm tissue culture dishes at 37˚C in an atmosphere containing 50 ml CO₂/l.

**Preparation of tumor lysate.** Four weeks after tumor inoculation, we resected the primary tumor lesion and soaked the entire tumor in liquid nitrogen to kill the tumor cells. The freeze-thawed tumor lysate was added to the DC cultures on Day 6 at a ratio of 5 DC equivalents to 1 tumor cell (i.e., 5:1). The homogenate was passed through a 0.2-µm filter to remove bacteria and tissues and mixed with the DCs for 24 h. After 24 h of incubation, nonadherent cells including DCs were harvested by gentle pipetting.

**Antibody.** The monoclonal antibody CTLA-4 (hamster IgG, mCD152 antibody; 0.2 mg/mouse) was provided by BioXcell (Lebanon, NH, USA). The CTLA-4 blocking activity was confirmed using the mink lung cell assay. The monoclonal control antibody IgG (hamster IgG, isotype control antibodies; 0.2 mg/mouse) was provided by BioXcell.

**Flow cytometry.** The markers Foxp3 and CD4, which are expressed on the surface of Tregs, were counted with a FACSCalibur™ Flow Cytometer (Becton Dickinson, San Jose, CA, USA) and were stained with fluorochrome-conjugated...
Antibody (BD Pharmingen, Tokyo, Japan) for the following markers: phycoerythrin (PE)-conjugated anti-mouse Foxp3 staining kit (eBioscience, San Diego, CA, USA) and fluorescein isothiocyanate (FITC)-conjugated rat anti-mouse CD4 (BD Pharmingen; clone: RM4-5). Data analysis was performed with CellQuest™ software (Becton Dickinson).

Immunofluorescence. Immunohistochemistry was used to measure the levels of Foxp3, a marker of Tregs, and CD8, a marker of cytotoxic T lymphocytes, within metastatic tumor lesions. Lung specimens were fixed in 20% formalin and embedded in paraffin. In each case, we examined all formalin-fixed, paraffin-embedded tumor tissue blocks. Five samples/mouse were cut into 4-µm slices. Rehydrated tissue sections were incubated with primary Abs against CD8+ (Santa Cruz Biotechnology, Santa Cruz, CA, USA) and Foxp3 (Abcam, Cambridge, MA, USA) diluted at 1:200 in Ab diluent (Dako ChemMate, Dako, Japan) overnight at room temperature. For CD8+ staining with FITC donkey anti-rabbit IgG and Foxp3+ staining with Texas red goat anti-rat IgG (Invitrogen, Carlsbad, CA, USA), secondary antibodies were diluted at 1:300 in Ab diluent (Dako ChemMate, Dako, Japan) overnight at room temperature. For CD8+ staining with FITC donkey anti-rabbit IgG and Foxp3+ staining with Texas red goat anti-rat IgG (Invitrogen, Carlsbad, CA, USA), secondary antibodies were diluted at 1:300 in Ab diluent and added for 60 min at room temperature in the dark. Digital images were captured using a BIOREVO microscope equipped with a confocal microscopy system (BZ-9000; Keyence, Japan).

Tumor volumes were measured using the micro-CT apparatus (R_mCT) to obtain high-resolution CT images in small living animals. The I-view-R (J. Morita Mfg Corp., Kyoto, Japan) was used as the viewer, and diagnosis was made with slice images viewed in all directions.

Tumor volumes were estimated using the formula (π x long axis x short axis x short axis)/6.

ELISA. We measured murine interferon-γ (IFN-γ) and interleukin-10 (IL-10) release by enzyme-linked immunosorbent assay using Quantikine® (R&D Systems, Minneapolis, MN, USA) according to the manufacturer’s protocol using an Easy Reader EAR340 microtest plate reader (SLT-Lab Instruments, Salzburg, Austria).

Statistical analysis. We determined differences among the 4 groups using a non-repeated measures analysis of variance (ANOVA) and the Scheffe test. All analyses were conducted using SPSS® 18.0 software (SPSS Japan, Inc., Tokyo, Japan). Results were expressed as the means ± standard deviation and P<0.01 was considered to indicate a statistically significant difference. For survival analysis, the differences in survival rates were analyzed by the log-rank test.

Results

Infiltration of CD8+ T lymphocytes and Tregs inside the tumor. The levels of Foxp3 were significantly decreased and the numbers of CD8+ cells were significantly increased inside the metastatic tumor lesions in the anti-CTLA-4 antibody-treated groups. Foxp3+ cells were not recruited to the metastatic area in the anti-CTLA-4 antibody-treated groups compared with the findings in the control IgG-treated group (Fig. 2A-D). The number of CD8+ T lymphocytes/unit area was higher (P<0.01) in the mice that received tumor lysate-pulsed DCs and the anti-CTLA-4 antibody (32.79±6.39 cells/mm²) compared to
those that received tumor lysate-pulsed DCs (16.98±5.16 cells/mm²) or anti-CTLA-4 antibody alone (17.41±2.88 cells/mm²; Fig. 2E). The number of Foxp3⁺ T lymphocytes/unit area was lower (P<0.01) in the mice that received the anti-CTLA-4 antibody (14.94±4.59 cells/mm²) compared to those that received tumor lysate-pulsed DCs (29.54±4.22 cells/mm²). The number of Foxp3⁺ T lymphocytes/unit area was lower (P=0.01) in the mice that received tumor lysate-pulsed DCs and the anti-CTLA4 antibody (6.29±6.12 cells/mm²) compared to those that received the anti-CTLA-4 antibody alone (Fig. 2F).

**Tumor volume of the lung metastases.** Forty-two days after inoculation, the volume of the metastatic lesion (P<0.01) in the mice that received tumor lysate-pulsed DCs and the anti-CTLA-4 antibody (319.58±49.96 mm³) was lower compared to those that received tumor lysate-pulsed DCs (669.04±40.19 mm³) or the anti-CTLA-4 antibody alone (611.54±31.97 mm³) (Fig. 3).

**Survival rate.** The median survival time was: IgG control, 32.27 days (range, 28-38); tumor lysate-pulsed DC-treated group, 43.47 days (range, 33-57); anti-CTLA4 antibody-treated group, 45.93 days (range, 39-60) and for the tumor lysate-pulsed DCs alone and anti-CTLA4 antibody alone groups were sacrificed within 42 days following formation of an enlarged tumor at the inoculated site. Survival was significantly prolonged but differences in the tumor lysate-pulsed DCs alone and the anti-CTLA4 antibody alone groups were small compared with the control IgG group (P<0.001). There was no significant difference between the tumor lysate-pulsed DCs alone and anti-CTLA4 antibody alone groups. Further lifetime prolongation was observed in the tumor lysate-pulsed group.
Discussion

Most osteosarcoma patients are treated with a combination of surgery and chemotherapy. Despite recent advances in local therapy with curative intent, chemotherapeutic treatments for metastatic disease often remain unsatisfactory owing to severe adverse effects and incomplete long-term remission. Therefore, the evaluation of novel therapeutic options is of great interest. Since the discovery of the regression of metastases after cryotreatment, investigators have examined immune responses in animal models in hopes of showing a ‘cryoimmunologic’ effect (12,23-25). Despite the lack of response to cryotreatment alone, we observed a synergistic effect when cryotreatment was combined with DCs (11). Since the initial discovery that CTLA-4 stimulation drives T-cell immunity (13-15), anti-CTLA-4 therapy has been used extensively for tumor immunotherapy. In addition, CTLA-4 stimulation has been demonstrated to suppress the function of Treg cells and drive potent CD8+ T cell-mediated tumor protection (16-19). However, the synergistic effect of anti-CTLA-4 antibody and cryotreated tumor lysate-pulsed DCs has not been investigated in osteosarcoma models. In the present study, we examined the synergistic effect of cryotreated tumor lysate-pulsed DCs and CTLA-4 blockade on preventing the development of a secondary tumor from a large primary tumor by using a mouse metastatic osteosarcoma model.

The anti-CTLA-4 antibody inhibited the accumulation of Tregs and induced the infiltration of CD8+ T cells inside the metastatic lesions. CTLA-4 signaling in CD4+Foxp3+ Tregs is required for their immunosuppressive capacity (13,16). Our findings revealed that CTLA-4 stimulation led to the inhibition of Foxp3+ T cells inside the tumor tissues. We report that after combination therapy, the numbers of intratumoral CD8+ T cells were significantly increased and Treg cells were depleted by the combination treatment, supporting the ability of the therapy to enhance the tumor-specific cell mediated immune response.

The group treated with the combination of tumor lysate-pulsed DCs and the anti-CTLA-4 antibody also displayed smaller lung metastases with a prolonged lifetime. Tregs comprise one of the major components of the immunosuppressive condition of tumor lesions (15). Noteworthy, the result of tumor rejection in lung metastatic lesions in the combined therapy group correlated with the intratumor ratio of CD8+ T cells to Tregs. This suggests that controlling immunosuppressive factors may facilitate the activity of DCs and cytotoxic T lymphocytes in the tumor. Identically, the Treg depletion using anti-CTLA-4 antibody treatment combined with tumor lysate-pulsed DCs treatment displayed a significantly improved survival in comparison to the tumor lysate-pulsed DCs or anti-CTLA-4 antibody monotherapy groups.

The combination therapy also resulted in the enhancement of the number of CD8+ T cells and prevented the proliferation of Tregs in the spleen, which is evidence of a systemic response with the potential to eradicate disseminated disease. Inhibition of Treg accumulation in the spleen enhances systemic cell-mediated immunity through the activation of DCs or cytotoxic T lymphocytes.

Tumor lysate-pulsed DCs and CTLA-4 blockade induced the activation of cell-mediated immunity by increasing serum IFN-γ levels and decreasing serum IL-10 levels. Tregs are among the major elements that cause potent immunosuppression mediated by cytokines from tumor cells and the inhibition of CTLA-4 stimulation may be useful for enhancing the efficacy of cancer therapy or vaccines (26-28). Our results revealed that blocking CTLA-4 signaling using anti-CTLA-4 antibody enhanced cell-mediated immunity.

Although the immune response to tumor lysate-pulsed DCs or anti-CTLA-4 antibody alone may vary or may be insufficient to suppress metastatic tumors, the combination of tumor lysate-pulsed DCs and CTLA-4 blockade has the potential to create a robust antitumor immune response that controls the growth of metastases. The present study could lead to a generation of proposals for clinical trials, in which tumor lysate-pulsed DCs will be combined with CTLA-4 blockade to treat cancer. Ipilimumab is a human monoclonal immunoglobulin G (IgG) antibody against CTLA-4, an immune inhibitory molecule that the US Food and Drug Administration has approved for the treatment of unresectable or metastatic melanoma in 2011 (29). The combination of ipilimumab and chemotherapy in the treatment of melanoma (30) and lung cancer (31) has been evaluated in clinical trials.

Chemotherapy for human osteosarcoma is the standard treatment and it should be performed before immunotherapy. However, additional methods have yet to be developed for treating patients who are resistant to the standard osteosarcoma treatment (32). Therefore, the development of novel treatment strategies for metastatic osteosarcoma is critical. The combination of chemotherapy, DC vaccination and anti-CTLA-4 antibody treatment may be effective in the treatment of osteosarcoma patients with residual tumors or distant metastasis after chemotherapy. These issues will be addressed in the future to enable the clinical application of our therapy in
treating osteosarcoma and to facilitate further research efforts.

Acknowledgements

We thank Hiroyuki Tsuchiya, Hideji Nishida, Katsuhiro Hayashi, Akihiko Takeuchi and Katsuro Tomita for participating in this study.

References


