Interleukin-4 receptor α-based hybrid peptide effectively induces antitumor activity in head and neck squamous cell carcinoma

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Abstract. Interleukin-4 receptor α (IL-4Rα) is highly expressed on the surface of various human solid tumors including head and neck squamous cell carcinoma (HNSCC). We designed a novel IL-4Rα-lytic hybrid peptide composed of a binding peptide to IL-4Rα and a cell-lytic peptide. In the present study, we evaluated the antitumor activity of the IL-4Rα-lytic hybrid peptide as a novel molecular-targeted therapy in HNSCC. Immunoblot analysis revealed that IL-4Rα was expressed in all tested HNSCC cell lines (HSC-2, HSC-3, HSC-4, Ca9-22 and OSC-19), but not in a human normal keratinocyte (HaCaT) cell line. Immunohistochemical expression levels of IL-4Rα in HNSCC tissues were higher compared to those in normal epithelial tissue. The IL-4Rα-lytic hybrid peptide showed cytotoxic activity in all five cancer cell lines with a concentration that killed 50% of all cells (IC₅₀) as low as 10 µM. HaCaT cells were less sensitive to this peptide with an IC₅₀ of >30 µM. In addition, intratumoral administration of IL-4Rα-lytic hybrid peptide significantly inhibited tumor growth in a xenograft model of human HNSCC in vivo. These results indicate that the IL-4Rα-lytic hybrid peptide may serve as a potent agent to provide a novel therapy for patients with HNSCC.

Introduction

Head and neck squamous cell carcinoma (HNSCC) which includes cancers of the oral cavity, oropharynx, larynx and hypopharynx is the sixth most common cancer worldwide and has an incidence of approximately 600,000 cases per year (1). The current management and treatment of HNSCC involves multi-modality approaches of surgery, chemotherapy and radiotherapy (2). Despite recent advances in early detection, diagnosis and treatment, the 5-year survival for patients with HNSCC has remained at 50% for the past 30 years (3). Molecular-targeted therapy, based on molecular findings of the last 50 years, is one of the most promising gateways to the development of new strategies in oncology (4). Cetuximab, a monoclonal antibody to the epidermal growth factor receptor (EGFR), is the only molecular-targeted therapy to be routinely used in clinical practice for the treatment of recurrent and metastatic HNSCC (5). Based on a limited number of phase II and III trials that have investigated the efficacy of cetuximab in addition to cisplatin in patients who were refractory to platinum-based therapy, the combination appears to confer further benefit over anti-EGFR agents alone (6,7).

The first generation of immunotoxins developed 35 years ago, which heralded targeted therapy, employed chemical conjugations of antibodies and either intact toxins or toxins with attenuated cell-binding properties. We previously reported that recombinant fusion protein IL4 (38-37)-PE38KDEL (also termed IL4-PE), consisting of circularly permuted interleukin (IL)-4 and a mutated form of Pseudomonas exotoxin (PE), induced significant regression of established biliary tract tumors and significantly improved the survival of animals with disseminated tumors (8,9). In addition, IL4-PE was reported to be highly and specifically cytotoxic to glioma cell lines in vitro, and caused partial or complete regression of established human glioblastoma multiforme tumors in nude mice (10). IL-4 receptor α (IL-4Rα)-targeted protein-based immunotoxin was tested in the clinic for the treatment of human solid tumors (11,12). However, its clinical application faced many challenges, including non-specific toxicities and immunogenicity (13).

To overcome these issues, we previously developed a ‘hybrid peptide’, composed of target-binding and cytotoxic
sequences containing cationic-rich D- and L-amino acids to form amphipathic partial α-helices that disrupt the cancer cell membrane selectively, and are stable when combined with a cancer-targeting moiety (14). It is known that peptide drugs are relatively easily synthesized using either recombinant or solid-phase chemical synthesis techniques and the production costs are generally affordable when compared to antibody-based therapeutics (14). IL-4Rα has been previously reported to have high expression on the surface of a variety of human solid tumors such as renal cell carcinoma, malignant melanoma and glioblastoma (15). Although the biological function of IL-4Rα expression on solid tumors remains unclear, this receptor may be an effective candidate for a novel molecular-targeted therapy.

In the present study, we examined the expression levels of IL-4Rα in both patient samples and HNSCC cell lines, and then explored the antitumor activity of the IL-4Rα-lytic hybrid peptide against HNSCC.

**Materials and methods**

**Patient samples.** HNSCC specimens were obtained from 5 patients (4 males, 1 female; mean age, 56.6 years), who underwent radical surgery at the Department of Oral and Maxillofacial Surgery, Tsukuba University Hospital, Japan from 2010 to 2011. Primary tumor sites were the tongue and the gingiva. For immunoblot analysis, we obtained HNSCC tissue from the cancerous lesion (cancer) and normal tissue from the normal area (normal) in other specimens from 1 patient. For immunohistochemistry, tissue was fixed in 10% formalin and paraffin embedded. The study protocol was approved in accordance with the ethics guidelines of the Tsukuba University (H23-61). All patients provided written informed consent for use of specimens.

**Peptides.** The following IL-4Rα-lytic hybrid peptide, the lytic peptide and IL-4 binding peptide were purchased from Invitrogen: the IL-4Rα-lytic hybrid peptide, KQLIRFLKLDRNGGGKLLKLLKLLKLKKK (underlined letters are D-amino acids); the lytic peptide, KLLKKLKKLLKLLKK; and the IL-4 binding peptide, KQLIRFLKRLDRN. All peptides were synthesized by the use of solid-phase chemistry, purified to homogeneity by reverse-phase high-pressure liquid chromatography and assessed by mass spectrometry. All peptides were dissolved in water and buffered to pH 7.4.

**Cell lines and culture conditions.** The HNSCC cell lines (HSC-2, HSC-3, HSC-4, Ca9-22 and OSC-19) were purchased from the Japanese Collection of Research Bioresources (Osaka, Japan). The human normal keratinocyte cell line (HaCaT) was purchased from the American Type Culture Collection (Manassas, VA, USA). These cell lines were maintained in Dulbecco's modified Eagle's medium or RPMI-1640 containing 10% heat-inactivated fetal calf serum (Nichirei Biosciences Inc., Tokyo, Japan) and 1% penicillin-streptomycin in a humidified atmosphere with 5% CO₂ at 37°C.

**Immunoblot analysis.** Immunoblot analysis was carried out as previously described (9). Briefly, whole-cell extracts were obtained using buffer containing 1% (v/v) Triton X-100, 0.1% (w/v) SDS, and 0.5% (w/v) sodium deoxycholate, separated by SDS-PAGE, and transferred onto a PVDF membrane. IL-4Rα antibody was used at dilution 1:200 (Santa Cruz Biotechnology, Santa Cruz, CA, USA) and actin as the internal control (Sigma-Aldrich, St. Louis, MO, USA). Proteins were visualized on Hyperfilm using an enhanced chemiluminescence/western blotting system (GE Healthcare, Piscataway, NJ, USA).

**Quantitative real-time PCR.** Total RNA of cells was isolated using High Pure RNA Tissue kit (Roche, Basel, Switzerland). For the reverse transcriptase reaction, 400 ng of the RNA sample was used. The reaction was carried out in a final volume of 10 µl of reaction mixture with Takara Kit II (Takara, Shiga, Japan). Aliquots (2 µl) of the cDNA samples were amplified in a final volume of 20 µl of PCR mixture containing SYBR Premix Ex Taq II (Takara). Quantitative real-time PCR was carried out using PRISM 7000 (Applied Biosystems, Carlsbad, CA, USA). The following primers were used: IL-4Rα forward, 5'-CTGACCTGGAGCAACCCGTATC-3' and IL-4Rα reverse, 5'-GCAGACGGACAAACCGTACAG-3'; GAPDH forward, 5'-GTCTTCACCACCATGGAAAGGCT-3' and GAPDH reverse, 5'-CATGCCAGTGAGCTTCCCGTTCA-3'.

**Cell viability assay.** Cell viability assay was performed as previously described (14). Briefly, cells were seeded into 96-well plates at 3x10⁴ cells/well in 90 µl of medium and incubated at 37°C for 24 h. Each peptide (IL-4Rα-lytic hybrid peptide, lytic peptide or IL-4 binding peptide) diluted in 10 µl culture medium was added to the cells. After a 72-h incubation, the cell viability assay using WST-8 solution (Nacalai Tesque, Kyoto, Japan) was performed.

**Immunohistochemistry.** For immunostaining of IL-4Rα, 2-µm sections from patient samples were stained using the Vectastain kit according to the manufacturer's instructions with the anti-IL-4Rα antibody (R&D Systems, Minneapolis, MN, USA).

**Binding assay.** The IL-4 binding peptide labeled with fluorescein isothiocyanate (FITC) was incubated with HSC-2 and HaCaT cells. Quantification of the binding activity of this peptide to HSC-2 and HaCaT cells treated with various concentrations for 30 min was carried out. Cells were washed twice with phosphate-buffered saline, and the peptides were detected using a flow cytometer (FACSCalibur, BD Biosciences, San Jose, CA, USA).

**Antitumor activity of the IL-4Rα-lytic hybrid peptide in a human tumor xenograft mouse model in vivo.** Animal experiments were carried out in accordance with the guidelines of Tsukuba University. HSC-2 cells (5x10⁶) resuspended in 150 µl of phosphate-buffered saline were inoculated subcutaneously into the flank region of 4- to 6-week-old athymic female nude mice weighing 17-20 g. When the tumors reached 20-60 mm³ in volume, animals were assigned randomly to four groups. Saline (control), the IL-4Rα-lytic hybrid peptide (5 or 10 mg/kg), or the lytic peptide alone (5 mg/kg) was injected intratumorally (50 µl/injection) three times a week for a total of 9 times. Tumors were measured with a caliper, and the tumor
Expression of IL-4Rα in HNSCC tissue specimens. We first analyzed the expression levels of IL-4Rα in HNSCC tissue specimens. Patient characteristics including age, gender, primary site of tumor, TNM classification and differentiation are shown in Table I. Immunoblot analysis and immunohistochemistry were performed to investigate the expression levels of IL-4Rα in tissue specimens from HNSCC patients. Immunoblot analysis showed that IL-4Rα was expressed in all HNSCC specimens (tongue and gingival carcinomas) but not in the normal tissue specimens from the same patients (Fig. 1A). Similarly, immunohistochemical analysis using anti-IL-4Rα antibody showed IL-4Rα immunopositivity in the HNSCC cancerous epithelium but not in the normal epithelium in all patients. Fig. 1B-E shows the staining pattern for patient no. 2.

Expression of IL-4Rα in cultured HNSCC cell lines. We next investigated the expression levels of IL-4Rα in HNSCC cell lines by immunoblot and real-time PCR analyses. Immunoblot and real-time PCR analyses demonstrated that all HNSCC cell lines expressed IL-4Rα but HaCaT did not (Fig. 2A). We also examined mRNA expression levels of IL-4Rα by real-time PCR analysis (Fig. 2B). Relative expression levels of IL-4Rα in HSC-2 and HSC-3 cells were ~4-fold higher than that of HaCaT cells. The lowest expression level of IL-4Rα in HNSCC cell lines was found in OSC-19, however, this level was still 2-fold higher than that of HaCaT (Fig. 2B).
Cytotoxic activity of the IL-4Rα-lytic hybrid peptide in HNSCC cell lines. To assess the in vitro cytotoxic activity of the IL-4Rα-lytic hybrid peptide in HNSCC and HaCaT cells, the WST assay was performed using HNSCC cell lines treated with the IL-4Rα-lytic hybrid peptide, lytic peptide or IL-4 binding peptide. HSC-2, HSC-3, HSC-4 and Ca9-22 cells were sensitive to the IL-4Rα-lytic hybrid peptide; the concentration that killed 50% of all cells (IC₅₀) was <5 µM. The OSC-19 cell line was also sensitive to the IL-4Rα-lytic hybrid peptide with an IC₅₀ of <10 µM. In contrast, optimal cell killing was not induced in HaCaT cells by either the lytic peptide or IL-4 binding peptide or IL-4Rα-lytic hybrid peptide (Fig. 3). The cytotoxic activity of the hybrid peptide was strongly enhanced when compared with that of the lytic peptide. The cytotoxic activity of the IL-4Rα-lytic hybrid peptide increased 4.0- to 13.2-fold when compared with that of the lytic peptide in the
We examined the binding activity of the IL-4 binding peptide labeled with FITC to both HSC-2 and HaCaT cells by flow cytometry, and then found that exposure of HSC-2 to this peptide resulted in the increased binding activity of this peptide in a concentration-dependent manner (Fig. 4). These results suggest that the IL-4Rα-lytic hybrid peptide selectively kills cancer cells expressing IL-4Rα.

In vivo antitumor activity of the IL-4Rα-lytic hybrid peptide in a human HNSCC xenograft mouse model. Following the observation that the IL-4Rα-lytic hybrid peptide exhibits a marked cytotoxic effect on HNSCC cells in vitro (Fig. 3), the antitumor activity of the hybrid peptide was assessed in a xenograft model of human HNSCC. HSC-2 cells were inoculated subcutaneously into athymic nude mice, and the animals were subsequently treated with the IL-4Rα-lytic hybrid peptide by intratumoral injection. As shown in Fig. 5, tumors grew aggressively in the control mice injected with saline alone, reaching a volume of >1000 mm³ by day 24. In contrast, mice treated with the IL-4Rα-lytic hybrid peptide showed significant tumor regression at both dosages: mean tumor volumes were 491 mm³ (5 mg/kg) and 283 mm³ (10 mg/kg) on day 24. Moreover, tumors in mice injected with the lytic peptide grew rapidly similar to tumor growth in the control mice with saline alone, reaching a volume of 1104 mm³ (Fig. 5). No other abnormalities, such as loss of appetite and body weight, were observed in mice injected with the IL-4Rα-lytic hybrid peptide (data not shown). Histological analysis also showed no side effects in tissues from the major organs, including the liver and kidney, which were obtained from mice treated with

Table II. Cytotoxic activity of each peptide in HNSCC and HaCaT cell lines.

<table>
<thead>
<tr>
<th>Cell lines</th>
<th>IC₅₀ (µM)</th>
<th>IC₅₀ ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td>HNSCC cells</td>
<td>IL-4Rα-lytic hybrid peptide</td>
<td>Lytic peptide</td>
</tr>
<tr>
<td>HSC-2</td>
<td>1.9±0.1</td>
<td>8.2±0.2</td>
</tr>
<tr>
<td>HSC-3</td>
<td>2.3±0.6</td>
<td>17.4±1.2</td>
</tr>
<tr>
<td>HSC-4</td>
<td>2.8±0.8</td>
<td>33.2±6.1</td>
</tr>
<tr>
<td>Ca9-22</td>
<td>2.2±0.7</td>
<td>29.6±5.8</td>
</tr>
<tr>
<td>OSC-19</td>
<td>8.2±5.4</td>
<td>33.3±13.1</td>
</tr>
</tbody>
</table>

IC₅₀ values (peptide concentration inducing 50% inhibition of control cell growth) are represented as means ± SD from triplicate determinations.

Figure 4. Binding assay of HSC-2 and HaCaT cells treated with the FITC-labeled IL-4 binding peptide. Quantification of the binding activity of this peptide in HSC-2 and HaCaT cells treated with various concentrations for 30 min. IL-4 binding peptide at 0 µM represents untreated cells used as a control. Fold-change in fluorescence intensity is the extent of binding of the FITC-labeled IL-4 binding peptide to each cell line, relative to the fluorescence intensity values. Data are presented as means ± SD (error bars) from triplicate determinations.

Figure 5. Antitumor activity of the IL-4Rα-lytic hybrid peptide in a xenograft model. (A) HSC-2 cells were inoculated subcutaneously into nude Balb/c mice. Animals received an intratumoral peptide injection or saline alone [ ], control; lytic peptide (○, 5 mg/kg or ▲, 10 mg/kg) three times per week from day 3 as indicated by the arrows. Data are expressed as mean ± SD (bars), n=6 animals per group. *P<0.01. (B-E) Images show tumor growth on day 24 after tumor inoculation: (B) saline alone; (C) lytic peptide; (D) IL-4Rα-lytic hybrid peptide (5 mg/kg); (E) IL-4Rα-lytic hybrid peptide (10 mg/kg).
intratumoral administration of the IL-4Rα-lytic hybrid peptide (data not shown). These results demonstrated that the IL-4Rα-lytic hybrid peptide exhibited effective antitumor activity in a mouse xenograft model of HNSCC.

Discussion

The main treatment options for patients with HNSCC currently involve surgery, radiotherapy and chemotherapy, alone or in combination. Despite significant advances in HNSCC treatment, survival rates and prognosis have improved only moderately over the years (16). Systemic chemotherapy remains the only effective treatment option, but it is associated with significant rates of toxicity in HNSCC patients, who usually have a high prevalence of co-morbidities and problematic lifestyle habits (17). Ideally, future therapies should act over the short term, to minimize damage to healthy cells and target tumor compartments that have the highest sensitivity.

The concept of a ‘magic bullet’ proposed by Paul Ehrlich over 100 years ago has led to the search for agents that can selectively target cancer cells (18). Immunotoxins are proteins used to treat cancer and are composed of an antibody fragment linked to a toxin (19). Several disadvantages of these conventional immunotoxins for clinical use include immunogenicity, undesirable toxicity, manufacturing difficulties, short half-lives and neutralizing antibody production (20,21). However, peptides can be produced affordably by chemical synthesis, with a cost comparable to that of producing protein drugs. Moreover, since peptides are easy to produce, a wide variety of candidate peptides combining moieties for targeting and for toxicity can be tested in preclinical settings. We previously linked two functional peptide domains to produce a novel chimerical peptide termed a ‘hybrid peptide’, which was designed as a bifunctional peptide that binds to receptors or proteins overexpressing in cancer cells and consequently disrupts the cancer membrane (14,22,23). In the present study, we focused on IL-4Rα as recent evidence suggests that IL-4Rα is preferentially expressed on the surface of a variety of solid tumors including HNSCC (24).

The high degree of antitumor activity of the IL-4Rα-lytic hybrid peptide in HNSCC correlated with the expression of IL-4Rα in vitro. All HNSCC tumor specimens showed specific immunohistochemical staining for IL-4Rα, and western blot analysis revealed expression of IL-4Rα. However, IL-4Rα expression was not observed in normal tissue specimens from the same patients (Fig. 1). These data are consistent with previous reports that HNSCC cells express IL-4Rα on their cell surface and confirm that IL-4Rα is expressed in situ (25,26). These results also indicate that this receptor may be an attractive target for the treatment of HNSCC.

Previous results suggest that IL-4 receptor-targeted cytotoxin may provide an effective therapeutic option for HNSCC (24,26). In the present study, the in vitro cytotoxicity of the IL-4Rα-lytic hybrid peptide was examined in five HNSCC cell lines (Fig. 3). HSC-2 cells, which showed the highest level of IL-4Rα expression in western blot analysis, also showed the highest sensitivity to the IL-4Rα-lytic hybrid peptide (Figs. 2 and 3). Normal HaCaT cells with low IL-4Rα expression were not sensitive to the IL-4Rα-lytic hybrid peptide (Fig. 3). These results suggest that the cytotoxic effect of the IL-4Rα-lytic hybrid peptide correlates well with the level of IL-4Rα expression.

In the present study, although the growth rate of HSC-2 was rapid, it was found that intratumoral administration of the hybrid peptide at 10 mg/kg, dramatically inhibited the growth of HSC-2 tumors in vivo (Fig. 5). Histological analysis also showed no abnormal changes in the tissues of major organs obtained from the mice injected with the hybrid peptide (data not shown). For clinical use, local injection may be effective for tumors such as HNSCC, and combination with prior chemoradiotherapy should be developed. These observations indicate that abundant IL-4Rα expression in HNSCC tumors would facilitate efficient targeting by the IL-4Rα-lytic hybrid peptide.

In conclusion, IL-4Rα was overexpressed in both tumor specimens from patients with HNSCC and in HNSCC cell lines in vitro. The overexpressed IL-4Rα on HNSCC cells could be successfully targeted with the IL-4Rα-lytic hybrid peptide in vitro and in vivo. Future investigations using cancer progenitor cells isolated from primary malignant tissues at different stages during cancer progression and metastatic disease may help to identify new biomarkers for the development of more effective diagnostic and prognostic methods and targeted therapies (27). Additional studies should be performed to reveal the antitumor activity of the IL-4Rα-lytic hybrid peptide in animal models, and perhaps a phase I clinical trial should be undertaken to study its antitumor activity.

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References


