Abstract. The association of TMEM45A with various cancers has been recently reported. However, the biological function of TMEM45A in ovarian cancer remains unclear. The present study aimed to elucidate the role of TMEM45A in regulating the biological behavior of ovarian cancer cells. We compared the expression of TMEM45A between ovarian cancer tissues and normal tissues based on RNA-sequencing data of the ovarian cancer cohort from The Cancer Genome Atlas (TCGA) project and our real-time PCR data from 25 pairs of ovarian cancer and their matched non-cancerous tissue samples. The expression of TMEM45A was then suppressed in two ovarian cancer cell lines, HO-8910 and A2780, by RNA interference. Cell proliferation, cell cycle distribution, adhesion and invasive ability were then detected using the Cell Counting Kit-8 assay (CCK-8), propidium iodide (PI) staining, and cell adhesion and Transwell assays, respectively. In addition, the mRNA and protein levels of transforming growth factor-β (TGF-β1 and TGF-β2), Ras homolog family member A (RhoA) and Rho-associated kinase 2 (ROCK2) were detected with real-time PCR and western blotting, respectively. TCGA data and our real-time PCR results demonstrated the overexpression of TMEM45A in ovarian cancer. Silencing of TMEM45A significantly inhibited cell proliferation and significantly increased the cell population in the G1 phase. Moreover, knockdown of TMEM45A also inhibited cell adhesion as well as cell invasion. More importantly, suppression of TMEM45A notably downregulated the expression of TGF-β1, TGF-β2, RhoA and ROCK2. In conclusion, TMEM45A may function as an oncogene for ovarian cancer, and inhibition of TMEM45A may be a therapeutic strategy for ovarian cancer.

Introduction

Ovarian cancer is the leading cause of mortality from gynecologic malignancies in the world (1). Ovarian cancer is often diagnosed in advanced stages, resulting in a poor survival rate (2). The 5-year survival rate of patients with advanced-stage disease is merely about 5-30% (3). Recent investigations to characterize genetic alterations in ovarian cancer have discovered extensive cytogenetic and molecular alterations in these tumors (4-9). However, only limited knowledge has been obtained regarding the basic molecular mechanisms that deregulate the growth of the ovarian epithelium and lead to the invasive and metastatic behavior of these tumors.

TMEM45A (also called DERP7, DNAPTP4 or FLJ10134) belongs to the large family of genes encoding predicted transmembrane (TMEM) proteins. Recently, there are a few studies concerning the expression and functions of TMEM45A in cancers (10-13). TMEM45A was reported as an epigenetically regulated gene in MCF-7 breast cancer cells (14). Overexpression of TMEM45A has been shown to favor chemoresistance of liver and breast cancer cells under hypoxic conditions (11). It has been reported that high expression of TMEM45A is associated with poor prognosis in patients with breast (11), bladder (12) and ovarian cancer (10). However, there remains a lack of in-depth research on the expression pattern and biological functions of THEM45A in ovarian cancers.

To investigate the role of TMEM45A in ovarian cancer, we compared its expression between ovarian cancer and normal tissues. The effects of TMEM45A silencing on the proliferation, adhesion and invasion of ovarian cancer cells were then assessed. The possible involved mechanisms were also explored. Our study provides evidence that TMEM45A is overexpressed in ovarian cancer and it may be an effective therapeutic target for this disease.

Materials and methods

Bioinformatic analysis. The Cancer Genome Atlas (TCGA) RNA-sequencing and corresponding clinical data were
downloaded from the TCGA website https://tcga-data.nci.nih.gov/tcga/ following approval of this project by the consortium. Data from 568 ovarian cancers and 8 adjacent normal tissues were used for RNA-seq analysis. To further investigate the biological pathways involved in ovarian cancer pathogenesis through the TMEM45A pathway, we performed a gene set enrichment analysis (GSEA) (15) by using GSEA version 2.0 from the Broad Institute at MIT. The data in question were analyzed in terms of their differential enrichment in a predefined biological set of genes (representing pathways). In this study, GSEA firstly generated an ordered list of all genes according to their correlation with TMEM45A expression, and then a predefined gene set (signature of gene expression upon perturbation of certain cancer-related gene) receives an enrichment score (ES), which is a measure of statistical evidence rejecting the null hypothesis that its members are randomly distributed in the ordered list. The expression level of TMEM45A was used as a phenotype label, and ‘metric for ranking genes’ was set to Pearson correlation. All other basic evidence rejecting the null hypothesis that its members are

Cancer specimens. Tumor tissues and paired non-cancerous tissues were collected from 25 patients diagnosed with epithelial ovarian serous adenocarcinoma, who were admitted to the Department of Gynecology and Obstetrics, Yangpu Hospital, Tongji University School of Medicine (Shanghai, China) between 2010 and 2013. Informed consent was obtained from all patients. This study was approved by the Ethics Committee of Yangpu Hospital, Shanghai, China.

Cell lines. OVCAR3, A2780, HO-8910, CAOV3, SK-OV-3 and HEK293T cells were obtained from the American Type Culture Collection (ATCC; Rockville, MD, USA). All culture media were supplemented with 10% fetal bovine serum (FBS), 100 mg/ml penicillin G, and 50 µg/ml streptomycin (all from Invitrogen Life Technologies, Carlsbad, CA, USA). The ovarian cancer cell lines, OVCAR3, A2780 and HO-8910, were cultured in RPMI-1640, and the CAOV3, SK-OV-3 and HEK293T cells were cultured in Dulbecco’s modified Eagle’s medium (DMEM) (all from Invitrogen Life Technologies). All cells were maintained at 37°C in 5% CO₂.

RNA extraction and real-time PCR. Total RNA was extracted using TRIzol reagent (Invitrogen Life Technologies) according to the manufacturer’s instructions. mRNA contained in 2 µg total RNA was reverse transcribed using a cDNA synthesis kit (Thermo Fisher Scientific, Rockford, IL, USA) according to the manufacturer’s instructions. Real-time PCR was performed to detect mRNA levels of the indicated genes. GAPDH served as an internal control. The primers used were: TMEM45A, 5'-AGGCCCTCATCACGAGAGTT-3' and 5'-CTTTCC AGAGGCTCCATCAGTGGT-3'; ROCK2, 5'-CATGCAATGTTAGTGTA AGGCTAAAGG-3' and 5'-GTAAGAGCGGTATCTTTAT-3'; ROCK1, 5'-GACTACTACGCCAAGGAGGTC-3' and 5'-GAG CTGGTTGTCATACTTC-3'; TGF-β1, 5'-GAGGCCTTTATCTTC AGT-3'. All reactions were conducted on an ABI 7300 real-time PCR machine (Applied Biosystems, Foster City, CA, USA) using the following cycling parameters: 95°C for 10 min, followed by 40 cycles of 95°C for 15 sec, and 60°C for 45 sec. The gene expression was calculated using the ΔΔCt method. All data represent the average of 3 replicates.

Western blotting. The cells were lysed with radioimmunoprecipitation assay buffer (50 mmol/l Tris-HCl, 150 mmol/l NaCl, 1% Triton X-100, 0.1% SDS and 1% deoxycholic acid sodium). The lysates were quantified using the BCA protein assay kit (Thermo Fisher Scientific). The lysates with equal amounts of protein were separated on SDS-PAGE gels followed by electrophoretic transfer to nitrocellulose membranes. Western blotting was then carried out with appropriate primary and horseradish peroxidase-conjugated secondary antibodies. Membranes were developed with enhanced chemiluminescence (Bio-Rad, Richmond, CA, USA). Antibodies against TMEM45A, TGFβ1, TGFβ2, RhoA and ROCK2 were purchased from Abcam (Cambridge, MA, USA). GAPDH antibody was from Cell Signaling Technology Inc. (Danvers, MA, USA).

Cell proliferation assay. A total of 3x10³ cells/well were plated in 96-well plates before viral infection and cultured for 24 h in normal conditions. They were then infected with the TMEM45A-shRNA virus or control virus (NC). Cell proliferation was detected at the indicated times using CCK-8 (Dojindo Laboratories, Kumamoto, Japan) according to the manufacturer’s instructions. Briefly, at the indicated time-points, CCK-8 solution (10 µl in 100 µl DMEM) was added to each well and incubated for 1 h. Optical density (OD) values at wavelength 450 nm were measured by a microplate reader (Bio-Rad Laboratories Inc., Hercules, CA, USA).

Cell cycle analysis. The cell cycle was evaluated by flow cytometry using propidium iodide (PI; Sigma, St. Louis, MO, USA) staining on a flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA). Briefly, the cells were plated in 6-well plates before viral infection and cultured for 24 h under normal conditions. The cells were collected and fixed in 70% ethanol at -20°C overnight 24 h after viral infection. The
cells were then washed in phosphate-buffered saline (PBS) and resuspended in staining solution containing 20 µg/ml PI and 200 µg/ml RNase A. Experiments were performed in triplicate and 3x10^4 cells were analyzed per sample. G1, S, and G2/M fractions were quantified using CellQuest software (BD Biosciences) and manual gating.

**Cell adhesion assay.** The adhesion assay was performed in 12-well plates. The plates were pre-coated with 1 ml of fibronectin (5 µg/ml) for 2 h at room temperature. The cells were infected with the indicated viral vectors 48 h before the assay was performed. The cells were seeded into the coated plates at a density of 10^5 cells/well and allowed to adhere at 37˚C for 1 h. Non-adherent cells were washed off with PBS and fixed in 4% paraformaldehyde and stained with Giemsa solution. The number of adherent cells was determined as described previously (16).

**In vitro invasion assay.** The upper well of the Transwell (Corning Inc., Corning, NY, USA) was coated with Matrigel (BD Biosciences, San Jose, CA, USA) at 37˚C in a 5% CO₂ incubator for 1 h. The indicated cells were serum-starved for 24 h, and then 500 µl of cell suspension containing 10^5 cells/ml were placed in the upper compartment of the chamber. Culture medium supplemented with 10% FBS (750 µl) was added into the lower well of the chamber. The plates were incubated for 48 h. At the end of the incubation, the cells on the upper surface of the filter were completely removed by wiping with a cotton swab. The cells that migrated into the lower well were washed with PBS, fixed in 4% paraformaldehyde and stained by 0.2% crystal violet. The invading cells were observed under a microscope (magnification, x100). Cells were counted in the central field of triplicate membranes.

**Statistical analysis.** The data were analyzed using the two-tailed Student’s t-test to calculate the statistical significance of difference between groups. The results are presented as the mean value ± SD. Statistically significant differences were defined as having a P-value <0.05.

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**Results**

**Overexpression of TMEM45A in ovarian cancer.** To explore the expression of TMEM45A in ovarian cancer, we compared TMEM45A by analyzing high throughput RNA-sequencing data of the ovarian cancer cohort from TCGA. TMEM45A expression was significantly higher in the ovarian cancer tissues than that in the adjacent tissues of the patients, which indicated that TMEM45A may be an oncogene in ovarian cancer (Fig. 1A).

To further confirm the overexpression of TMEM45A in ovarian cancer, we performed real-time PCR analysis on 25 pairs of ovarian cancer and their matched non-cancerous tissue samples. TMEM45A was found to be overexpressed in 84% (21/25) of the tested ovarian cancer tissues (Fig. 1B). Statistical analysis using the Student’s t-test indicated a significant difference in the mRNA level of TMEM45A between the ovarian tumor and normal tissues (P<0.001).

**Downregulation of TMEM45A by RNA interference (RNAi) in ovarian cancer cells.** The protein and mRNA levels of TMEM45A in five ovarian cancer cell lines were then detected. A high level of TMEM45A was observed in the HO-8910 and A2780 cells (Fig. 2A). For that reason, these two cells were selected for the following assays.

To explore the functions of TMEM45A in ovarian cancer, we suppressed TMEM45A levels in the HO-8910 and A2780 cells by lentiviral-mediated delivery of shRNA. In the present study, three pairs of shRNA sequences against the human TMEM45A gene were designed and a non-specific scramble shRNA sequence was used as the negative control (NC). The silencing effect of the TMEM45A-RNAi virus was confirmed in the HO-8910 and A2780 cells (Fig. 2A). For that reason, these two cells were selected for the following assays.

**TMEM45A knockdown inhibits cell proliferation and induces G1-phase cell cycle arrest in ovarian cancer cells.**
To investigate the role of TMEM45A downregulation on cell proliferation, we assessed the proliferation of HO-8910 and A2780 cells infected with TMEM45A-Ri-3 using the CCK-8 assay. As shown in Fig. 2D and E, cell growth was notably impaired in the TMEM45A-Ri-3 viral-infected cells (TMEM45A-Ri-3) compared to the wild-type (WT) cells and the scramble shRNA viral-infected (NC) cells. Our data indicated a role of TMEM45A in the regulation of ovarian cancer cell proliferation.

The possible effect of TMEM45A RNAi on cell cycle progression was then determined. As shown in Fig. 2F and G, a higher number of HO-8910 cells in the G1 phase (70.8±1.9%) was observed in cells infected with the TMEM45A-shRNA-3 viral-infected cells, compared with that in the WT cells (45.1±1.1%) and the scramble shRNA viral-infected cells. The percentages of cells in the G1, S and G2-M phases for each sample at 48 h after viral infection are shown. The data are shown as the mean value ± SD. **P<0.01, vs. NC.
Similar results were observed in the NC cells (45.9±1.1%). Similar results were observed in the A2780 cells (Fig. 2G). These results indicated that the proliferation-promoting function of TMEM45A was most likely mediated by promoting G1/S cell cycle transition in the ovarian cancer cells.

Suppression of TMEM45A expression represses cell adhesion capacity. The effects of TMEM45A on cell adhesion capacity were evaluated by a cell adhesion assay (Fig. 3). The adherent ability of ovarian cells to fibronectin was significantly repressed by TMEM45A knockdown. The number of adherent TMEM45A-Ri-3 cells was 37% of that of the NC cells when the HO-8910 cells were used (Fig. 3A and C). Similar results were obtained for the A2780 cells (Fig. 3B and D). These data suggest that TMEM45A may regulate ovarian cancer cell adhesion.

TMEM45A knockdown inhibits the invasiveness of ovarian cancer cells. To discover whether TMEM45A affects the invasive ability of ovarian cancer cells, we performed a Matrigel-coated membrane chamber invasion assay. As shown in Fig. 3, a radical reduction in the invasive ability was observed in the TMEM45A-knockdown cells compared to the control cells. The number of invaded TMEM45A-Ri-3 cells decreased to 38% of that of the NC cells when the HO-8910 cells were used (Fig. 3E and G). Similar results were observed for the A2780 cells (Fig. 3F and H).

Identification of genes and signaling-associated biological pathways and processes by GSEA. The exact pathway that TMEM45 may regulate in ovarian cancers remains unclear. To probe the TMEM45A-associated pathways on an unbiased basis, we performed GSEA using high throughput RNA-sequencing data of the ovarian cancer cohort from TCGA. GSEA is designed to detect coordinated differences in expression of predefined sets of functionally related genes. Among all the 188 predefined ‘KEGG pathway’ gene sets, the TGF-β signaling pathway and focal adhesion pathway were identified as having a significant association with higher expression of TMEM45A (Fig. 4A and B).

TMEM45A knockdown downregulates the mRNA and protein levels of TGF-β1, TGF-β2, RhoA and ROCK2. To further confirm the association of TMEM45A expression and the TGF-β signaling and focal adhesion pathways, we detected the expression of important regulators of the TGF-β signaling
pathway and cell adhesion molecules in the TMEM45A-Ri-3 and NC cells. The mRNA and protein levels of TGF-β1, TGF-β2, RhoA and ROCK2 were significantly decreased in both the HO-8910 (Fig. 4C) and A2780 (Fig. 4D) cells following downregulation of TMEM45A.

**Discussion**

Recently, the expression and functions of TMEM45A in cancers have been reported (10-13). In the present study, bioinformatic analysis using high throughput RNA-sequencing data from TCGA ovarian cancer demonstrated a higher expression of TMEM45A in ovarian cancer compared to that in normal tissues (Fig. 1A). We then evaluated the mRNA levels of TMEM45A in 25 pairs of ovarian tumor and normal tissues by real-time PCR. We found that TMEM45A was overexpressed in 84% (21/25) of the tested ovarian cancer tissues (Fig. 1B).

To further investigate the functions of TMEM45A in ovarian cancer, we suppressed the expression of TMEM45A in two ovarian cell lines, HO-8910 and A2780, by RNA interference (Fig. 2B and C). Our data showed that suppression of TMEM45A expression markedly repressed the cell proliferation (Fig. 2D and E). Abnormal regulation of the cell cycle is frequently observed in most common malignancies, resulting in aberrant cell proliferation (17,18). In the present study, silencing of TMEM45A by RNAi significantly stimulated cell cycle arrest in the G1-phase (Fig. 2F and G), which indicated that TMEM45A promoted cell proliferation by promoting G1/S cell cycle transition in ovarian cancer cells. Moreover, cell adhesion and invasion of ovarian cells were also inhibited by TMEM45A downregulation (Fig. 3). These data suggest a role of TMEM45A in ovarian cell carcinogenesis.

The exact pathway that TMEM45 may regulate in ovarian cancers remains unclear. Our GSEA data indicated that TMEM45A overexpression was positively correlated with the TGF-β signaling pathway (Fig. 4A). TGF-β signaling participates in a variety of cellular processes, including cell differentiation, proliferation, apoptosis, and determination of developmental fate (19). The TGF-β signaling pathway has also been considered to promote tumor progression and
invasion. In response to elevated TGF-β levels, the tumor cell becomes more migratory and invasive (20,21). In the present study, TMEM45A knockdown significantly decreased the expression of TGF-β1 and TGF-β2 (Fig. 4), which indicates an association between TMEM45A function and the regulation of TGF-β signaling in ovarian cancer cells.

Moreover, our GSEA results indicated a positive correlation between TMEM45A overexpression and focal adhesion genes (Fig. 4B). Rho is required for the formation and maintenance of focal adhesion. It has been shown that RhoA, a member of the Rho subfamily, plays a central role in the regulation of cell survival, motility, apoptosis and invasion (22,23). RhoA expression was reported to be upregulated in lung, breast, colon and ovarian cancer. The expression level of RhoA may be positively correlated with the progression of these carcinomas, suggesting that RhoA may play an important role in tumorigenesis and tumor progression (24-28). The malignant phenotype in gastric cancer (29) and breast cancer (30) cells can be reversed by the inhibition of RhoA expression. Suppression of ROCK2 expression was reported to impair anchorage-independent growth and invasion of non-small cell lung cancer (31). In the present study, TMEM45A knockdown markedly decreased the mRNA and protein levels of RhoA and ROCK2. The downregulation of RhoA and ROCK2 may be associated with the impaired adhesion and invasive ability (Fig. 3) of the TMEM45A RNAi cells. To our knowledge, our data are the first to associate the functions of TMEM45A with the RhoA/ROCK2 signaling pathway. In conclusion, our study demonstrated the overexpression of TMEM45A in ovarian cancer, which was related with cancerous transformation. Our study is the first to link TMEM45A with the regulation of TGF-β signaling and the RhoA/ROCK signaling pathway, and thus provides useful information for targeted therapy.

References