miR-1246 and miR-4644 in salivary exosome as potential biomarkers for pancreatobiliary tract cancer

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Abstract. Pancreatobiliary tract cancer is a highly fatal cancer. Detection of pancreatobiliary tract cancer is difficult because it lacks typical clinical symptoms and because of its anatomical location. Biomarker discovery is therefore important to detect pancreatobiliary tract cancer in its early stage. A study demonstrated that expression levels of miR-1246, miR-3976, miR-4306, and miR-4644 in serum exosomes were higher in pancreatic cancer patients than these levels in healthy control participants. Supposing that microRNA (miRNA) expression profiles in saliva are similar to those in serum, four miRNAs (miR-1246, miR-3976, miR-4306, and miR-4644) in salivary exosomes may also be useful for detection of pancreatobiliary tract cancer. In this study, it was examined whether these miRNAs could be used as biomarkers for pancreatobiliary tract cancer. Twelve pancreatobiliary tract cancer patients and 13 healthy control participants were analyzed as a cancer and a control group, respectively. Unstimulated whole saliva was collected, salivary exosomes were isolated, and total RNA was extracted. Using quantitative real-time PCR (RT-qPCR), the relative expression ratios of miR-1246 and miR-4644 were significantly higher in the cancer group than these ratios in the control group. Receiver operating characteristic (ROC) curves were constructed to analyze the discrimination power of these miRNAs. For miR-1246, the results yielded an area under the curve (AUC) of 0.814 (P=0.008). For miR-4644, the results yielded an AUC of 0.763 (P=0.026). For the combination of miR-1246 and miR-4644, the results yielded an increased AUC of 0.833 (P=0.005). This pilot study suggests that miR-1246 and miR-4644 in salivary exosomes could be candidate biomarkers for pancreatobiliary tract cancer.

Introduction

Pancreatobiliary tract cancer is a term used to describe malignant carcinoma in pancreatic, gallbladder, and extrahepatic bile ducts. Pancreatobiliary tract cancer is a highly fatal cancer (1-4). Detection of pancreatobiliary tract cancer is difficult because it lacks typical clinical symptoms and because of its anatomical location, especially in the early stage. Biomarker discovery is therefore important for identification of increased risk, early diagnosis, and prediction of response to therapy and prognosis of pancreatobiliary tract cancer (5,6). The serum carbohydrate antigen 19-9 (CA19-9) level is one clinically useful biomarker related to pancreatobiliary tract cancer (6-8). However, there is also still a limit to detect pancreatobiliary tract cancer using the existing methods (9). Therefore, further studies are needed to develop biomarkers for pancreatobiliary tract cancer.

Recently, microRNAs (miRNAs) have been reported as potential biomarkers for various types of cancers including pancreatobiliary tract cancer (10-15). They are single-stranded non-coding RNAs of ~21-23 nucleotides in length that can bind to the three prime untranslated region (3'‑UTR) of target messenger RNAs (16). It has been noted that miRNAs could be detected in body fluids (17). Several studies have suggested that miRNAs circulating in blood are useful as biomarkers for the diagnosis of pancreatic cancer (10-14,18).

Recent studies have also focused on salivary miRNAs as biomarkers for various diseases (19). Saliva collection is simple and non-invasive. Saliva contains proteins, nucleic acids, and hormones originating from both local and systemic
sources. Notably, salivary miRNAs have been shown to be highly stable because exosomes or protein complexes protect them (20). A few studies have suggested that salivary miRNAs are useful as biomarkers for the diagnosis of pancreatic cancer (21,22). However, in these studies, exosomes were not extracted from saliva. Most miRNAs in human saliva likely exist in exosomes (23,24). For diagnosis of pancreatobiliary tract cancer, miRNAs in salivary exosomes may be superior to those in saliva.

To the best of our knowledge, there is no literature regarding the relationship between miRNAs in salivary exosomes and pancreatobiliary tract cancer. On the other hand, one report is available regarding the relationship between miRNAs in blood exosomes and pancreatic cancer (18). In that study, four miRNAs (miR-1246, miR-3976, miR-4306, and miR-4644) in serum exosomes were identified as biomarkers for pancreatic cancer. miRNA expression profiles in saliva could be similar to those in serum (25). Therefore, in the present study, we hypothesized that miR-1246, miR-3976, miR-4306, and miR-4644 in salivary exosomes might be useful biomarkers for pancreatobiliary tract cancer. Thus, the aim of this study was to examine whether these four miRNAs in salivary exosomes could be useful biomarkers for pancreatobiliary tract cancer.

Materials and methods

Study population. The study design was a case-control study. It was not possible to estimate the sample size preliminarily, because there was no prior information on which to base a sample size (26).

Twelve patients (6 males and 6 females) with pancreatobiliary tract cancers were referred to the Department of Preventive Dentistry, Okayama University Hospital for saliva collection in the morning before the onset of cancer therapy from July 2013 to July 2014.

As a control group, 13 healthy participants were recruited at the Department of Preventive Dentistry, Okayama University Hospital from August 2014 to December 2014 for saliva collection in the morning. Inclusion criteria for healthy control participants were ≥50 years of age and no history of any cancer. To avoid the effect of systemic conditions on circulating miRNA expressions, the exclusion criteria for healthy control participants were as follows: diabetes (27); pulmonary diseases (28); cardiovascular diseases (29); kidney diseases (30); liver diseases (31); and autoimmune diseases (32) at the time of saliva collection. According to the inclusion/exclusion criteria, the control participants with hypertension and dyslipidemia (2 participants) and hyperuricemia (1 participant) were included in this study. This study was approved by the Ethics Committee, Okayama University Graduate School of Medicine, Dentistry and Pharmaceutical Sciences and Okayama University Hospital (no. 1506-052). Written, informed consent was obtained from all participants.

General status examination. Measurements were performed before the onset of cancer therapy. Medical charts were reviewed to obtain information about cancer and the body mass index (BMI). A personal interview was performed to gather information about smoking habits (pack-years). Serum levels of hemoglobin A1c (HbA1c), C-reactive protein (CRP), albumin, carcinoembryonic antigen (CEA), and CA19-9 were evaluated. Concentrations of HbA1c, CRP, and albumin were measured by the high-performance liquid chromatography method, the latex agglutination method, and the bromocresol green albumin method, respectively. Concentrations of CEA and CA19-9 were measured by electrochemiluminescence immunoassays.

Saliva collection. Unstimulated whole saliva was collected as reported previously with minor modification (24). Briefly, saliva was collected in the morning (7:00 a.m.-12:00 noon). During the collection period, participants were seated straight up and instructed to refrain from speaking or swallowing. They allowed the saliva to accumulate in the floor of the mouth and then spit it through a funnel into a tube kept on ice. At least 0.5 ml of unstimulated whole saliva was collected. After collection of saliva, it was stored at 4°C for up to 6 h, after which it was stored at -80°C until use.

Exosome isolation. Exosomes were isolated from saliva samples (0.5-1.0 ml) using Total Exosome Isolation Reagent (Invitrogen, Carlsbad, CA, USA), in accordance with the manufacturer's instructions (33).

RNA extraction. Total Exosome RNA and Protein Isolation Kits (Invitrogen) were used for extraction of total RNA from exosome samples. Total RNA was extracted in accordance with the manufacturer's instructions. After extraction of total RNA from salivary exosomes, their quality was confirmed using the Agilent 2100 Bioanalyzer and the Agilent RNA 6000 Pico Kit (both from Agilent Technologies, Santa Clara, CA, USA).

Quantitative real-time PCR (RT-qPCR). To compare miRNA expression between the control and cancer group, TaqMan RT-qPCR assays were performed. TaqMan MicroRNA Assays (Life Technologies, Carlsbad, CA, USA) were used for the RT-qPCR analyses performed on the Mx3000P Real-Time QPCR System (Agilent Technologies) according to the manufacturer's instructions (34,35). Briefly, reverse transcription (RT) enzymes including specific RT primers for each miRNA and total RNA sample were mixed in a tube. RT run conditions were then set. The plate was incubated at 16°C for 30 min, 42°C for 30 min, and 85°C for 5 min, and then held at 4°C. Products of the RT reaction were stored at -80°C until use. RT-qPCR was performed in triplicate using TaqMan Universal Master Mix II, no UNG (Applied Biosystems/Life Technologies, Carlsbad, CA, USA). After holding at 95°C for 10 min, 50 thermal cycles (95°C for 15 sec and 60°C for 60 sec) were run. Threshold cycle (Ct) values were determined using the background-based threshold (cycle-range, 5-8) calculated by the MxPro Mx3000P v4.10 software (Stratagene/Agilent Technologies, Edinburgh, UK). Data with a raw Ct>40 for each miRNA were treated as Ct=40 for the subsequent statistical test (36). U6 snRNA was considered suitable as an internal control (22). The relative expression rates of each miRNA were calculated using U6 snRNA expression.

Discrimination power of candidate miRNA biomarkers for pancreatobiliary tract cancer. To evaluate the discrimination power of candidate miRNA biomarkers for pancreatobiliary tract cancer.
tract cancer, receiver operating characteristic (ROC) curves (37) were constructed. The ROC curves were obtained by plots of the sensitivity (true-positive rate) and 1-specificity (false-positive rate) of the diagnostic test at various cut-off values. The ROC curves graphically show the relationship between the true- and the false-positive rate according to the various cut-off values. The area under the curve (AUC) was estimated to assess the predictive power. To decide the optimal threshold value, the Youden index (sensitivity + specificity - 1) was used (38).

Statistical analyses. Characteristics of the control and cancer group are represented as continuous variables [age, BMI, smoking habit (pack-years), HbA1c, CRP, albumin, CEA, and CA19-9] and categorical variables (gender, cancer site, and cancer stage). The Mann-Whitney U test, the Chi-square test, and Fisher's exact test were used to assess significant differences in clinical variables between the two groups, as appropriate. The Mann-Whitney U test was used to compare the relative expression ratios of each miRNA of the control and cancer group. To calculate the P-value for the AUC, a non-parametric test for AUC=0.5 was performed. Two-sided P<0.05 values were considered to represent significant differences. To assess the correlations of variables in the cancer group, Spearman's rank correlation coefficient and its P-value were calculated for age (years), HbA1c (NGSP) (%), CRP (mg/dl), albumin (g/dl), CEA (ng/ml), CA19-9 (U/ml), relative expression ratios of the miRNAs, and smoking (pack-years) as continuous variables, and cancer stage (I-III, IVa, and IVb) as a categorical variable. These statistical analyses were performed using SPSS software version 20 (SPSS, Inc., Chicago, IL, USA).

Results

Characteristics of the participants. Table I summarizes the characteristics of all participants. There were no significant differences in age and gender between the two groups. However, a significant difference in the percent of current smokers was observed (P=0.015). Pancreatic cancer was predominant in the patient group (75.0%), and 83.3% of patients were diagnosed as having the most advanced (IVa or IVb) stage. In addition, 58.3 and 66.7% of patients showed values greater than the reference values of CEA (10 ng/ml) and CA19-9 (100 U/ml), respectively.

RT-qPCR validation. After the RT-qPCR assays, miR-3976 was excluded from the following analyses because 24 of 25 (96.0%) samples had high (>40) raw Ct-values. The number of samples having high (>40) raw Ct-values of expression levels of miR-1246, miR-4306, miR-4644, and U6 snRNA were 0, 2, 1 and 0, respectively. Therefore, miR-1246, miR-4306, and miR-4644 were considered as candidate miRNAs. For the control group, the median (25th and 75th percentile) values of log2-transformed relative expression of miR-1246, miR-4306, and miR-4644 were 11.6 (10.9, 12.6), -5.4 (-6.8, -3.9), and -6.5 (-7.6, -4.9), respectively. For the cancer group, those of miR-1246, miR-4306, and miR-4644 were 14.7 (12.6, 16.2), -5.0 (-8.1, -2.9), and -4.1 (-5.7, -2.2), respectively. The Mann-Whitney U test showed that the relative expression ratios of miR-1246 and miR-4644 were significantly higher in the cancer than these ratios in the control group (Fig. 1).

<table>
<thead>
<tr>
<th>Characteristics</th>
<th>Control (n=13)</th>
<th>Cancer (n=12)</th>
<th>P-valuea</th>
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</thead>
<tbody>
<tr>
<td>Age (years)</td>
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<tr>
<td>Median (range)</td>
<td>66 (53-83)</td>
<td>65 (45-84)</td>
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<td>Gender n (%)</td>
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<tr>
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<td>6 (46.2)</td>
<td>6 (50.0)</td>
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<td>Female</td>
<td>7 (53.8)</td>
<td>6 (50.0)</td>
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<tr>
<td>Smoking, n (%)</td>
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<tr>
<td>Current</td>
<td>0 (0.0)</td>
<td>5 (41.7)</td>
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<td>Cancer site, n (%)</td>
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<tr>
<td>Bile duct cancer</td>
<td>2 (16.7)</td>
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<tr>
<td>Gallbladder cancer</td>
<td>1 (8.3)</td>
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<tr>
<td>Pancreatic cancer</td>
<td>9 (75.0)</td>
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<td>T categoryb, n (%)</td>
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<tr>
<td>T1</td>
<td>0 (0.0)</td>
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<td>T2</td>
<td>1 (8.3)</td>
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<td>T3</td>
<td>2 (16.7)</td>
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<td>T4</td>
<td>7 (58.3)</td>
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<td>2 (16.7)</td>
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<td>N categoryb, n (%)</td>
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<tr>
<td>N0</td>
<td>6 (50.0)</td>
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<td>Cancer stageb, n (%)</td>
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<tr>
<td>I</td>
<td>0 (0.0)</td>
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<td>II</td>
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<td>III</td>
<td>1 (8.3)</td>
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<td>IVa</td>
<td>4 (33.3)</td>
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<tr>
<td>IVb</td>
<td>6 (50.0)</td>
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<td>HbA1c (NGSP) (%)</td>
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<tr>
<td>Median (range)</td>
<td>5.9 (4.9-8.1)</td>
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<tr>
<td>CRP (mg/dl)</td>
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<tr>
<td>Median (range)</td>
<td>0.42 (0.06-5.47)</td>
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<td>Albumin (g/dl)</td>
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<tr>
<td>Median (range)</td>
<td>3.7 (2.9-4.5)</td>
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<tr>
<td>CEA (ng/ml)</td>
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<tr>
<td>Median (range)</td>
<td>12 (1-1.379)</td>
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<tr>
<td>CA19-9 (U/ml)</td>
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<tr>
<td>Median (range)</td>
<td>414 (1-38.864)</td>
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*aMann-Whitney U test, Chi-square test, or Fisher's exact test was used.

General Rules for the Study of Pancreatic Cancer (Japan Pancreas Society) were used. HbA1c, hemoglobin A1c; CRP, C-reactive protein; CEA, carcinoembryonic antigen; CA19-9, carbohydrate antigen 19-9.
Discrimination power of miR-1246 and miR-4644 for pancreatobiliary tract cancer. To evaluate the discrimination power of miR-1246 and miR-4644 for pancreatobiliary tract cancer, ROC curves were constructed (Fig. 2). For miR-1246, the results yielded an AUC of 0.814 [95% confidence interval (CI), 0.616-1.000; cut-off, 13.77; sensitivity, 0.667; specificity, 1.000; P=0.008]. For miR-4644, the results yielded an AUC of 0.763 (95% CI, 0.564-0.961; cut-off, -5.205; sensitivity, 0.750; specificity, 0.769; P=0.026). For the combination of miR-1246 and miR-4644, the results yielded an increased AUC of 0.833 (95% CI, 0.630-1.000; cut-off, 8.035; sensitivity, 0.833; specificity, 0.923; P=0.005).

Correlations of variables in the cancer group. In the cancer group, there were significant correlations between CRP and cancer stage (r=0.822), between albumin and HbA1c (r=0.582), between albumin and CRP (r=0.711), between CA19-9 and miR-1246 (r=0.818), between miR-1246 and miR-4644 (r=0.671), between smoking and HbA1c (r=0.579), and between smoking and albumin (r=0.647) (Table II).

Discussion
This study was performed to clarify whether miR-1246, miR-3976, miR-4306, and miR-4644 levels in salivary...
Table II. Correlation of variables in the cancer group.

<table>
<thead>
<tr>
<th>Variables</th>
<th>Values</th>
<th>Age (years)</th>
<th>Cancer stage (I-III, IVa, and IVb)</th>
<th>HbA1c (NGSP) (%)</th>
<th>CRP (mg/dl)</th>
<th>Albumin (g/dl)</th>
<th>CEA (ng/ml)</th>
<th>CA19-9 (U/ml)</th>
<th>mirR-1246&lt;sup&gt;a&lt;/sup&gt;</th>
<th>mirR-4306&lt;sup&gt;a&lt;/sup&gt;</th>
<th>mirR-4644&lt;sup&gt;a&lt;/sup&gt;</th>
<th>Smoking (pack-years)</th>
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<tbody>
<tr>
<td>Age (years)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
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<tr>
<td>Cancer stage</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.652</td>
<td></td>
<td>0.483</td>
<td>0.111</td>
<td>0.111</td>
<td>0.326</td>
<td>0.031</td>
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<td>(I-III, IVa, and IVb)</td>
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<tr>
<td>HbA1c (NGSP) (%)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.111</td>
<td></td>
<td>0.822&lt;sup&gt;d&lt;/sup&gt;</td>
<td>0.626</td>
<td>0.514</td>
<td>0.001</td>
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<tr>
<td>CRP (mg/dl)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.326</td>
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<td>0.582&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.047</td>
<td>0.099</td>
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<td>Albumin (g/dl)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>0.527</td>
<td>0.078</td>
<td>0.340</td>
<td>0.078</td>
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<tr>
<td>CEA (ng/ml)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.541</td>
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<td>0.578</td>
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<tr>
<td>CA19-9 (U/ml)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>0.527</td>
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<td>0.179</td>
<td>0.578</td>
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<td>miR-1246&lt;sup&gt;a&lt;/sup&gt;</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
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<td>0.231</td>
<td>0.214</td>
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<td>miR-4306&lt;sup&gt;a&lt;/sup&gt;</td>
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<td>miR-4644&lt;sup&gt;a&lt;/sup&gt;</td>
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<td>0.812</td>
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<td>0.527</td>
<td>0.983</td>
<td>0.470</td>
<td>0.578</td>
<td>0.983</td>
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<tr>
<td>Smoking (pack-years)</td>
<td>r&lt;sup&gt;b&lt;/sup&gt; P-value&lt;sup&gt;b&lt;/sup&gt;</td>
<td>0.681</td>
<td></td>
<td>0.527</td>
<td>0.983</td>
<td>0.470</td>
<td>0.578</td>
<td>0.983</td>
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<sup>a</sup>Log2-transformed relative expression ratio of each microRNA using U6 snRNA as reference was analyzed. <sup>b</sup>Spearman's rank correlation coefficient and its P-value. <sup>c</sup>P< 0.05, <sup>d</sup>P< 0.01. HbA1c, hemoglobin A1c; CRP, C-reactive protein; CEA, carcinoembryonic antigen; CA19-9, carbohydrate antigen 19-9.
exosomes were useful as potential biomarkers for pancreato-
biary tract cancer. Among these four miRNAs, two miRNAs
(miR-1246 and miR-4644) in salivary exosomes showed
significantly higher expression in pancreatobiliary tract
cancer patients than that noted in healthy control participants.
In addition, in the ROC curve analysis, the AUCs of both
miR-1246 and miR-4644 were >0.7, indicating fair discrimi-
natory power (39). These results indicate that miR-1246 and
miR-4644 in salivary exosomes could be useful biomarkers
for screening pancreatobiliary tract cancer patients. On the
other hand, in the present study, overlap of the relative expres-
sion profiles of miR-1246 and miR-4644 was also found in
the cancer and control groups. This suggests that miR-1246
and miR-4644 might be non-specific screening biomarkers for
pancreatobiliary tract cancer.

Compared to other body fluids including blood, saliva
may be immediately exposed to the outside environment and
confounded by a wide variety of environmental factors (19).
However, some molecules in saliva are highly stable and
represent the body's health status. In this study using salivary
exosomes, the sensitivity and specificity for detecting pancreato-
biliary tract cancer were 66.7 and 100.0% for miR-1246 (cut-off,
13.77) and 75.0 and 76.9% for miR-4644 (cut-off, -5.205),
respectively. In serum, it is reported that sensitivity and spe-
cificity for pancreatic cancer of miR-1246 were 79.1 and 82.0%
respectively (40). It is also known that, for pancreatic cancer, in
plasma they were 82 and 73% for miR-885-5p, 82 and 82% for
miR-22-3p, and 82 and 55% for miR-642b-3p, respectively (11).
Taking these findings into account, the sensitivity for pancrea-
tobiliary tract cancer of miR-1246 and miR-4644 in salivary
exosomes seems to be slightly lower than other miRNAs in
blood, and their specificity seems to be similar or greater than
that of other miRNAs in blood.

In the present findings, the combination of miR-1246 and
miR-4644 in salivary exosomes increased the sensitivity for
pancreatobiliary tract cancer. This is consistent with previous
results, indicating that concomitant pancreatic cancer-initi-
ating cell and miRNA marker expression strengthened the
sensitivity for pancreatic cancer (18). In the present study,
whereas drawing blood from healthy participants was difficult
ethically, we could not combine miRNAs in salivary exosomes
and serum markers to assess the predictive power. However,
serum markers would further strengthen the sensitivity for
pancreatobiliary tract cancer of miR-1246 and miR-4644 in
salivary exosomes.

Serum CA19-9 is one of the most studied and validated
serum biomarkers for pancreatobiliary tract cancer (6-8). In
the present study, the serum CA19-9 level was significantly
correlated with the expression of miR-1246 and miR-4644 in the
serum CA19-9 was also found in the cancer and control groups.
This suggests that miR-1246 and miR-4644 might be non-specific screening biomarkers for
pancreatobiliary tract cancer.

In conclusion, the present results demonstrated that
miR-1246 and miR-4644 in salivary exosomes increased the sensitivity for
pancreatobiliary tract cancer. This is consistent with previous
results, indicating that concomitant pancreatic cancer-initi-
ating cell and miRNA marker expression strengthened the
sensitivity for pancreatic cancer (18). In the present study,
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pancreatobiliary tract cancer.

miR-1246 has been found to be aberrantly expressed
in pancreatic cancer tissue (41), and such a condition could
contribute to the increased miRNA in blood. Due to the exten-
sive blood supply in salivary glands, saliva is considered to
be a terminal product of blood circulation, and molecules that
are present in blood are also present in saliva. Therefore, it is
feasible that miR-1246 in salivary exosomes originated from
pancreatobiliary tract cancer tissue. However, further studies
are needed to clarify this point.

Studies have investigated the relationship between salivary
miRNA profiles and pancreatobiliary tract cancer. For instance,
a clinical study showed that miR-21, miR-23a, miR-23b, and
miR-29c were significantly upregulated in saliva of pancreatic
cancer patients compared to controls (21). Another clinical
study also reported that salivary miR-3679-5p was significantly
downregulated and miR-940 was significantly upregulated in
pancreatic cancer (22). Among the present findings, miR-1246
and miR-4644 in salivary exosomes were confirmed to show
significant cancer-related increases. The present and previous
findings support the notion that salivary miRNAs are useful
biomarkers for pancreatobiliary tract cancer. A saliva test that
is less invasive than a blood test or endoscopy may be used as
a broad screening test to identify pancreatobiliary tract cancer
patients who would need further screening.

The limitations of this study are as follows. First, external
validity was limited because all participants were recruited
at the Okayama University Hospital. In addition, the cancer
patients were mainly diagnosed as having advanced-stage
disease when they participated in this study. Further studies
including patients with chronic pancreatitis and early stage
of pancreatobiliary tract cancer are necessary to improve the
validity of miR-1246 and miR-4644 in salivary exosomes
as screening markers of pancreatobiliary tract cancer.
Furthermore, overall screening of miRNAs in salivary
exosomes using a microarray will be necessary to discover a
new biomarker in pancreatobiliary tract cancer patients.

In conclusion, the present results demonstrated that
miR-1246 and miR-4644 in salivary exosomes could be
useful biomarkers for identification of patients with pancrea-
tobiliary tract cancer. We hope that clinical use of simple and
non-invasive salivary tests will contribute to the screening
of pancreatobiliary tract cancer patients and improve their
survival.

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