Abstract. Squamous cell carcinoma of the lung is one of the most aggressive cancers, and its aggressiveness is in part due to its intrinsic high rate of metastasis. Moreover, the process of epithelial-mesenchymal transition (EMT) appears to be involved in these neoplastic processes. Furthermore, EMT-type cells share many biological characteristics with the function of angiogenin (ANG) in squamous cell lung carcinoma. We conducted immunohistochemical analysis to detect the expression of ANG, E-cadherin, vimentin, N-cadherin, β-catenin and TGF-β1 in 60 cases of squamous cell lung carcinoma tissues. Western blot analysis was adopted to detect the protein expression levels of ANG and EMT markers. The effects of ANG on proliferation, migration and invasion of squamous cell lung carcinoma cells was analyzed by Cell Counting Kit-8, scratch assay and Transwell invasion chamber in order to reveal the role of ANG in the process of EMT in squamous cell lung carcinoma. The results revealed that ANG was aberrantly expressed in the squamous cell lung carcinoma specimens and was closely correlated with the differentiation of the cell lines. The expression of ANG was also significantly associated with metastasis and the stage of the squamous cell lung carcinoma cases. In addition, we validated that ANG influenced the expression of vimentin, E-cadherin, N-cadherin, β-catenin and TGF-β1 in SK-MES-1 cells. Most importantly, overexpression of ANG enhanced the migration and invasion of SK-MES-1 cells, while knockdown resulted in opposite effects. In the present study, we found that ANG plays an important role in EMT in squamous cell lung carcinoma and may be a valuable therapeutic target for squamous cell lung carcinoma.

Introduction

With the advancements in research on mutations in the epidermal growth factor receptor (EGFR) kinase, as well as fusions involving anaplastic lymphoma kinase (ALK), the treatment of patients with lung adenocarcinoma has experienced marked improvement (1,2). Unfortunately, the therapeutic effect of these personalized treatment programs is less obvious in the second most common type of lung cancer, squamous cell lung carcinoma (SQCLC). Hence, it is necessary to identify other significant markers as new targets for SQCLC therapy.

Angiogenin (ANG), a 14,400 Da angiogenic ribonuclease of the RNase A superfamily, is upregulated in a variety of human cancers and has a 33% amino acid identity and an overall homology of 56% to that of RNase-A (3). Angiogenin was once thought to promote cancer progression through its angiogenic activity. However, recent research has demonstrated that ANG can directly stimulate tumor cell proliferation and its expression and activity were found to be upregulated significantly in a variety of human tumors including breast, colorectal, kidney and pancreatic cancers, and this upregulation was found to be correlated with tumor progression and poor prognosis in patients (4). As a transcription factor, ANG cannot be detected in normal, non-endothelial cells (5), however, its expression in human cancer cells and human tumors has not been widely investigated.

More importantly, ANG is implicated in a wide range of biological functions including cell survival, growth, proliferation, migration, tube formation and tumor angiogenesis (6-8). Epithelial-mesenchymal transition (EMT), a process in which epithelial cells lose or modify their apical-basal polarity and are converted to a mesenchymal phenotype, is a critical process during embryonic development and tumor metastasis (9,10). The process of EMT is typically characterized...
by losing the expression of the epithelial markers, E-cadherin and β-catenin, which are transmembrane proteins essential for stable adherens junctions, and acquiring the expression of mesenchymal markers, such as N-cadherin, vimentin and TGF-β (11,12). We, therefore, hypothesized that ANG is an inducer of EMT in human squamous cell lung carcinoma and enhances the metastatic potential.

Materials and methods

Tissue collection. The cancerous and corresponding non-tumor normal specimens of SQCLC, freshly frozen for western blot analysis and qRT-PCR, were obtained from 50 patients who underwent lung cancer resection procedures at Changhui Hospital from October, 2012 to October, 2015. All cancerous and matching non-cancerous samples used for this study were provided by the Clinical and Experimental Pathology of the Research Centre of Changhui Hospital, Shanghai, China where initial H&E staining and histologic diagnoses were performed after standard surgical primary tumor resection. This study was conducted in accordance with the Declaration of Helsinki and was approved by the Medical Ethics Committee of Changhui Hospital, with all patients providing written informed consent.

Cell culture. SK-MES-1 cells were obtained from the Cancer Cell Repository (Shanghai Cell Bank, Shanghai, China), and were maintained in Dulbecco’s modified Eagle’s medium (DMEM) supplemented with 10% heat-inactivated fetal bovine serum (Gibco-BRL) and antibiotics (100 U/ml penicillin and 100 U/ml streptomycin; Hyclone Laboratories, Inc., Logan, UT, USA) at 37°C in a humidified atmosphere of 5% CO₂.

RNA interference and ANG overexpression. The ANG-specific short hairpin RNA adenovirus vector (shANG) and the negative control short hairpin RNA adenovirus vector (shScramble) were purchased from Genechem Co., Ltd. (Shanghai, China). The sequences of shANG were: sense strand 1: 5' -CCCGCGG ATGACAGATCAGTGTGAGAGATCTACAGTCTGGTA CCCC-3’ and antisense strand 1: 5’-AACGGGATGACAGATACTGTGAAGAGATTCACAGTATCTGTCA-3’. The sequences of shANG were: sense strand 2: 5' -CCGGATCCCAGGCTCGTTCTTTGTCTCCTACAAAGAACGAGCCTGGGATCC-3’ and antisense strand 1: 5’-AACGGGATGACAGATACTGTGAAGAGATTCACAGTATCTGTCA-3’.

Quantitative real-time reverse-transcription polymerase chain reaction (qRT-PCR). Total RNA was extracted and cDNA was synthesized using the PrimeScript RT Reagent kit (Takara Bio). SYBR Premix Ex Taq (Takara Bio) was used for quantitative real-time polymerase chain reaction performed using a LightCycler® 480 Real-Time PCR system (Roche Diagnostics). The primers for each gene were as follows: 18S rRNA forward: 5’-CGGACACGGACAGGATTGCAC-3’; 18S rRNA reverse: 5’-GCATGGCAAGATHCTCTCGT-3’; ANG forward: 5’-GGACCTTGTTTCTGAGCCGAG-3’; ANG reverse: 5’-CCAGCACAGAAGCACAACA-3’. The amplification conditions were as follows: 1 cycle of 95°C for 30 sec and 40 cycles of 95°C for 5 sec followed by 60°C for 30 sec. The expression levels of the target genes relative to the control were determined using the 2^ΔΔCT method. 18S rRNA served as the internal control.

Western blot analysis. Western blot analyses were performed as previously described (13). The primary antibodies included: angiogenin (Sc-74528, Santa Cruz Biotechnology, Inc., 1:1,000), vimentin (ab92547, Abcam, 1:2,000), TGF-β1 (ab92486, Abcam, 1:1,000), E-cadherin (ab40772, Abcam, 1:10,000) and β-catenin (60008-1-Ig, Proteintech, 1:2,000).

Immunohistochemistry. Immunohistochemical assay was performed as previously described (14). The primary antibodies used were: angiogenin (Sc-74528, Santa Cruz Biotechnology, Inc., 1:1,000), vimentin (ab92547, Abcam, 1:200), TGF-β1 (ab92486, Abcam, 1:100), E-cadherin (ab40772, Abcam, 1:500), N-cadherin (ab12221, Abcam, 1:500), and β-catenin (ab32572, Abcam, 1:500).

Immunofluorescence. Paraformaldehyde-fixed SK-MES-1 cells were first incubated with primary antibodies at 4°C overnight. After being washed three times with PBS, the cells were incubated with secondary antibodies for 1 h at 37°C. Nuclei were counterstained with 4',6-diamidino-2-phenylindole (DAPI) for 2 min. The primary antibodies used were angiogenin (Sc-74528, Santa Cruz Biotechnology, Inc., 1:1,000), E-cadherin (ab40772, Abcam, 1:500) and N-cadherin (ab12221, Abcam, 1:500). The secondary antibodies used were Alexa Fluor 498-conjugated anti-rabbit IgG (1:400, Jackson ImmunoResearch).

Scratch assay. Cells were placed in a 24-well plate at an initial density of 1x10⁵ cells/well. A uniform monolayer formed overnight. After being washed three times with PBS, the cells were incubated with secondary antibodies for 1 h at 37°C. A pipette tip was used to create a wound in the monolayer by scraping. Images were captured at 24 h after the scratch and the migrated cells were quantitated.

Invasion assay. The invasion assay was performed using Transwell insert chambers with a pore size of 8 μm (Corning). The Transwell filter inserts were coated with Matrigel; 0.5x10⁵ cells were seeded in serum-free medium in the upper chamber. After a 24-h incubation at 37°C, cells in the upper chamber were carefully removed with a cotton swab and the cells that had traversed the membrane were fixed, stained in a 0.1% crystal violet solution and counted.

Cell proliferation. Cell proliferation was analyzed by using a Cell Counting Kit-8 (Beyotime, China). Twelve hours after being plated into a 96-well plate at a density of 2,000 cells/well the cells were transfected with an adenovirus. Next, the cells were incubated for 0, 24, 48 and 72 h; then, 10 µl CCK-8 solution was added to each well and the cultures were incubated at 37°C for 1 h. Subsequently, the absorbance at 450 nm was measured.

Statistical analysis. All statistical analyses were performed using SPSS version 19.0. The quantitative data were first evalu-
ated on whether they followed the normal distribution by the Shapiro-Wilk test. The data with a non-normal distribution were analyzed using the Kruskal-Wallis test. The data with a normal distribution were assessed using the Student's t-test. A P-value <0.05 was considered to be a statistically significant difference.

Results

ANG is expressed in SQCLC and positively correlates with a high grade of malignancy of SQCLC. Immunohistochemical assays showed that ANG was expressed in SQCLC tissues and is correlated with poor postoperative surgical-pathologic staging. (A) Immunohistochemical assays showing ANG expression in low-grade and high-grade SQCLC and normal lung tissues adjacent to tumors. (B) Percentage of normal and cancer cases with high, moderate and negative staining for ANG. (C) Percentage of low-grade and high-grade cases with high, low and negative staining for ANG. ANG, angiogenin; SQCLC, squamous cell lung carcinoma; LG, low-grade; HG, high-grade.

Figure 1. ANG is expressed in SQCLC tissues and is correlated with poor postoperative surgical-pathologic staging. (A) Immunohistochemical assays showing ANG expression in low-grade and high-grade SQCLC and normal lung tissues adjacent to tumors. (B) Percentage of normal and cancer cases with high, moderate and negative staining for ANG. (C) Percentage of low-grade and high-grade cases with high, low and negative staining for ANG. ANG, angiogenin; SQCLC, squamous cell lung carcinoma; LG, low-grade; HG, high-grade.

Figure 2. Upregulation of angiogenin (ANG) enhances the proliferation, migration and invasion of SK-MES-1 cells. (A) mRNA and protein levels of ANG were significantly enhanced in the Ad-ANG SK-MES-1 cells (n=3 for each cell group). Ad-Null served as the control. *P<0.05 vs. control. (B) Scratch assay was used to evaluate the migration capability of the SK-MES-1 cells in 2-dimensional space (magnification, x100). Ad-ANG SK-MES-1 cells showed a higher migration capability. Quantified data are presented as the number of migrated cells per high-power field (HPF) 24 h after scratch. Ad-Null served as the control. *P<0.05 vs. control. (C) Higher optical density at 450 nm in the CCK-8 assay was observed in the Ad-ANG SK-MES-1 cells at 24, 48 and 72 h, respectively. *P<0.01 vs. control. (D) Transwell invasion chamber assay was used to evaluate the invasion capability of the SK-MES-1 cells in 3-dimensional space (magnification, x200). Ad-ANG SK-MES-1 cells showed higher invasion capability. Quantified data are presented as the number of invasive cells per HPF. All experiments were carried out in triplicate. *P<0.01 vs. control.
tissues, while it was not detected in the normal lung tissues adjacent to tumors (Fig. 1A). As shown in Fig. 1B, ANG was significantly increased in the SQCLC tissues compared with that noted in the normal lung tissues (P<0.0001). Notably, high-grade SQCLC which indicates a higher postoperative surgical-pathologic staging exhibited the strongest ANG expression (Fig. 1A). Consistently, the difference in the patterns of ANG expression in low-grade and high-grade SQCLC was significant (low-grade vs. high-grade, P<0.0001, Fig. 1C), which showed that a high percentage of high-grade SQCLC cases had strong ANG expression when compared with the expression in its low-grade counterpart. Collectively, our results indicated that ANG expression increased with the malignant grade of SQCLC.

Upregulation of ANG enhances proliferation, migration and invasion of SK-MES-1 cells in vitro. Since the local expression level of ANG was considerably increased in the SQCLC tissues, especially in the cancerous tissues from high-grade SQCLC patients where ANG expression was intensely positive, as compared with the adjacent normal tissue, we considered that ANG may play an important role in the development, metastasis and invasiveness of SQCLC. To investigate the effect of ANG on SQCLC, we used an in vitro system. Firstly, we effectively infected SK-MES-1 cells with Ad-ANG (Fig. 2A). Evaluation of cell proliferation was performed after infection using a CCK-8 assay kit. It was found that Ad-ANG SK-MES-1 cells manifested increased proliferation levels at 24, 48 and 72 h, as compared with the shScramble SK-MES-1 cells (n=3 for each cell group, *P<0.05 vs. the control). (B) shANG SK-MES-1 cells showed a lower migration capability. Quantified data are presented as the number of migrated cells per high-power field (HPF) 24 h after being scratched. shScramble served as the control. (C) Ad-ANG SK-MES-1 cells showed a lower proliferation capacity at 24, 48, and 72 h, as compared with the shScramble SK-MES-1 cells (n=3 for each cell group, *P<0.05 vs. the control). (D) Transwell invasion chamber assay was used to evaluate invasion capability of the SK-MES-1 cells in 3-dimensional space (magnification, x200). shANG SK-MES-1 cells showed lower invasion capability. Quantified data are presented as the number of invasive cells per HPF. All experiments were carried out in triplicate. *P<0.05 vs. the control.

Figure 4. Immunohistochemical staining of mesenchymal and epithelial markers, including TGF-β1, vimentin, N-cadherin, E-cadherin and β-catenin in SQCLC. Compared with low-grade SQCLC, epithelial markers including E-cadherin and β-catenin were significantly reduced, while mesenchymal markers including N-cadherin, vimentin and TGF-β1 were markedly enhanced in the high-grade SQCLC. SQCLC, squamous cell lung carcinoma; LG, low-grade; HG, high-grade.
also invasion capability (Fig. 2B and D), as compared with the Ad-Null cells.

These results definitely indicated that ANG is an important regulator of proliferation, migration and invasion in SK-MES-1 cells.

Knockdown of ANG reduces the ability of proliferation, migration and invasion of SK-MES-1 cells in vitro. We further used shANG to knock down ANG in SK-MES-1 cells. The cells were infected with shANG and cultured for 48 h. SK-MES-1 cells infected with shScramble were used as a negative control. Compared with the shScramble SK-MES-1 cells, the expression level of ANG mRNA in the shANG cells was decreased by 70% (Fig. 3A). Western blot analysis results also confirmed lower expression level of ANG in the shANG SK-MES-1 cells (Fig. 3A). Furthermore, it was revealed that the shANG SK-MES-1 cells showed decreased proliferation capability (Fig. 3C). Meanwhile, decreased migration and invasion capacities were also observed in the shANG SK-MES-1 cells, as compared with the shScramble SK-MES-1 cells (Fig. 3B and D).

EMT is associated with the grade of malignancy of SQCLC. Given the fact that EMT contributes to metastasis and invasiveness of various cancers, we examined the expression of several EMT markers in both low-grade and high-grade SQCLC tissues. As shown in Fig. 4, compared with low-grade SQCLC, epithelial markers including E-cadherin and β-catenin were downregulated and the expression of mesenchymal markers including N-cadherin, vimentin and TGF-β1 were upregulated in high-grade SQCLC tissues, indicating that EMT may contribute to a metastatic and aggressive phenotype of SQCLC.

Overexpression of ANG upregulates the expression of mesenchymal markers but downregulates epithelial markers in the SK-MES-1 cell line. After showing that both ANG and EMT were positively correlated with a high grade of malignancy of SQCLC, we further tested whether ANG induces the metastatic and invasive phenotype of SQCLC by augmenting EMT. As shown in Fig. 5A, we found that the E-cadherin staining was decreased while N-cadherin staining was increased in the Ad-ANG SK-MES-1 cells as compared with the Ad-Null cells. Furthermore, western blot analysis confirmed that the expression of vimentin and TGF-β1 was significantly enhanced in the Ad-ANG cells. In contrast, Ad-ANG SK-MES-1 cells exhibited a lower level of protein expression of epithelial markers including E-cadherin and β-catenin, as compared with the Ad-Null and negative control groups (Fig. 5B and C, respectively; P<0.01). These data indicated that ANG triggers the process of EMT in SK-MES-1 cells.

Discussion

ANG, a 14.4 kDa polypeptide, was first isolated from serum-free supernatants of a human adenocarcinoma cell line, HT-29 (15). In addition to angiogenic activity, ANG is also implicated in a variety of biological functions including cell
growth, proliferation, migration and tube formation (6,8,16). Of note, some studies have reported that ANG can undergo nuclear translocation to stimulate RNA transcription in both cancer cells and cancer-associated endothelial cells (17). Recent research has shown that the expression and activity of ANG are upregulated in many human tumors including breast, kidney, colorectal and lung cancers (18). In the present study, immunohistochemical assays showed that ANG was expressed in SQCLC tissues, while it was not expressed in normal lung tissues adjacent to tumors, indicating that ANG is involved in the development of SQCLC, which is consistent with the finding that increased ANG is positively associated with the incidence of human tumors.

ANG has been reported as a member of the urokinase plasminogen activator receptor interactome that participates in the formation of plasmin and the migration of breast cancer cells, which are necessary for tumor metastasis and invasion (19). In a clinical study, researchers found that, among the markers representing different aspects of cancer vascular biology and exhibiting abnormal expression in colorectal cancer, ANG was the only index which was independently associated with the increasing stage of colorectal cancer according to the Dukes' and American Joint Committee on Cancer systems (20). Similarly, in patients with adenocarcinoma, our previous study showed that increased expression of ANG was correlated with vascular and pleural invasion as well as positive lymph node metastasis (6). Consistently, we found that increased expression of ANG was positively correlated with the high grade of SQCLC while knockdown of ANG abrogated proliferation, migration and invasion of SK-MES-1 cells. These results from clinical specimens and cultured squamous cell lung carcinoma cell lines supported the notion that ANG contributes to the invasive phenotype of squamous cell lung carcinoma cells.

Numerous studies have demonstrated that cell invasion during tumor progression may be dependent on the acquisition of EMT features (21). Evidence from clinical studies also suggests that poor survival of cancer patients and drug-resistance are associated with EMT phenotypes in malignant cancer cells (22). EMT is a complex process during which epithelial cells lose their polarity and cell-cell adhesion, exhibit enhanced cell-extracellular matrix adhesion, gain invasive properties and become mesenchymal-like cells. In both squamous cell lung carcinoma tissues and SK-MES-1 cells, we showed that high expression of ANG was positively correlated with the expression of mesenchymal markers but negatively associated with epithelial markers. These data suggest that ANG induces EMT in squamous cell lung carcinoma. The intermediate filament protein vimentin is an important marker of EMT and a critical regulator of mesenchymal cell migration. Induction of vimentin is associated with increased expression of Axl which enhanced the migratory activity of pre-malignant breast epithelial cells (23). Cell adhesion molecules are important to cancer invasion and metastasis. E-cadherin to N-cadherin switch was found to promote cancer progression via TGF-β-induced EMT in extrahepatic cholangiocarcinoma (24) and is of strong importance in the progression of prostate cancer (25). In lung cancer cell line, H1650ER, N-cadherin expression was also reported to be upregulated and paralleled by the reduced expression of E-cadherin. Proliferation and invasion of H1650ER cells were inhibited by knockdown of N-cadherin (26). In accordance with these previous studies, our results indicated that overexpression of ANG led to the upregulation of vimentin, fibronectin and N-cadherin and reduced expression of E-cadherin, suggesting that ANG contributes to invasion and metastasis of lung carcinoma by inducing EMT.

Many molecular and cell signaling pathways have been reported to participate in the EMT process. For instance, accumulating evidence indicates that the phosphatidylinositol 3-kinase (PI3K)/Akt/mTOR signaling pathway plays an important role in EMT (27,28). The MAPK/ERK pathway has also been demonstrated to be critical for many features of EMT including acquisition of invasive properties, enhanced MMP activity, and attenuation of adhesions. EMT mediated by TGF-β also involves the activation of the MAPK/ERK pathway (29). In addition, research has shown that receptor activator of NF-κB (RANK) can induce EMT in human mammary epithelial cells and promotes oncogenesis and metastasis (30). Notably, correlation of ANG and these cell signaling pathways has been demonstrated in many studies. Overexpression of ANG can activate phosphorylation of downstream molecules of the PI3K/AKT/mTOR signaling pathway in bladder cancer cells (31). In cultured human umbilical vein endothelial cells, ANG induced the activation of both ERK1/2 and SAPK/JNK (32,33). Moreover, ANG prevented stress-induced death of P19 embryonal carcinoma cells via upregulation of the Bcl-2 and NF-κB pathways (34). Given these results from extensive studies, we speculate that ANG may induce EMT via a complex regulating network involving various cell signaling pathways.

In summary, this study suggests that ANG promotes the invasion and metastasis of SQCLC by enhancing proliferative, migratory and invasive ability of squamous cell carcinoma cells through the induction of EMT. Our results highlight the possibility that ANG may serve as a target for the treatment of squamous cell carcinoma of the lung.

Acknowledgements

This study was supported by the National Natural Science Foundation of China (nos. 81272592 and 81301829).

References


