Abstract. Baicalein, a naturally occurring flavonoid isolated from the roots of *Scutellaria baicalensis*, is historically and widely used as anti-inflammatory and anticancer therapy. Nevertheless, the anti-metastatic effect and underlying molecular mechanisms of baicalein on colorectal carcinoma (CRC) remain unclear. The aim of the present study was, therefore, to investigate the anti-metastatic activity of baicalein and related mechanism(s) on CRC cells. In this study, we observed that baicalein treatment inhibited proliferation, as well as migration and invasion of HT-29 and DLD1 cells. Baicalein decreased the expression of the matrix metalloproteinases-2 (MMP-2) and MMP-9 in a dose-dependent manner. Also, baicalein treatment significantly reduced phosphorylation of extracellular signal regulated kinases (ERK). Furthermore, in DLD1 cells, MEK1 overexpression partially blocked the anti-metastatic effects of baicalein. Combined treatment with an ERK inhibitor (U0126) and baicalein led to the synergistic reduction of MMP-2/9 expression; and the invasive capabilities of DLD1 cells were also inhibited markedly. Finally, intragastric administration of baicalein inhibited CRC xenograft growth *in vivo* and suppressed the phosphorylation of ERK and the expression of MMP-2/9 in tumor tissues. Consequently, baicalein suppresses CRC cell invasion via inhibition of the ERK signaling pathways, indicating that baicalein is a potential agent for CRC treatment.

Introduction

CRC is one of the leading causes of death worldwide (1). Aggressive local invasion and metastasis make CRC difficult to treat (2). Therefore clarifying the mechanism of invasion of CRC to find new treatment targets is urgent.

Metastasis of cancer cells is a complex processes during which the degradation of the extracellular matrix (ECM) plays an important role (3). MMP-2/-9 play important roles in degrading ECM and its involvement is complex in the process of cancer metastasis (3,4). Overactivity of ERK signaling pathway has been found in almost half of known human tumor cell lines and in many human primary tumors derived from different origins (5). ERK signaling pathway plays a vital role in the synthesis of MMP-2/9 (3,6). Overactivity of ERK pathway is observed in CRC (7,8) and is correlated with the metastasis of CRC (9,10).

Baicalein is extracted from the roots of *Scutellaria baicalensis* or *Scutellaria radix*. The chemical structure of baicalein is shown in Fig. 1A. The antitumor biological effect of baicalein has been found in many tumors (11-13). Baicalein has been found to possess antitumor effect in hepatocellular carcinoma, glioma and bladder cancer (3,14-16), but the anti-metastatic effect and related mechanism(s) in CRC are still unclear. Thus, the present study investigated the effects of baicalein in CRC invasion and metastasis and related mechanisms.

Materials and methods

Cell lines and reagents. CRC HT29 and DLD1 cell lines were purchased from the American Type Culture Collection (ATCC; Manassas, VA, USA) and cultured in RPMI-1640 (HyClone Laboratories, Inc., Logan, UT, USA) medium with 10% fetal bovine serum (FBS; HyClone Laboratories). The cells were cultured at 37˚C with 5% CO₂. Anti-MMP-2, ERK, MMP-9 and p-ERK were purchased from Cell Signaling Technology (Beverly, MA, USA). Unless otherwise indicated, all the other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Apoptosis assays. Apoptotic and/or necrotic cells were analysed by propidium iodide (PI) uptake using Annexin V binding an Annexin V-FITC/PI kit as previously described (15). Briefly, CRC cells were seeded into 6-well plates at a density of 1x10⁵ cells/well for 12 h and treated with 0, 40, 60 and 80 μM of baicalein for 48 h. After washing with cold phosphate-buffered saline (PBS), cells were resuspended in Annexin V binding buffer. Cells were incubated at 37°C with 5% CO₂. Anti-MMP-2, ERK, MMP-9 and p-ERK were purchased from Cell Signaling Technology (Beverly, MA, USA). Unless otherwise indicated, all the other chemicals were purchased from Sigma-Aldrich (St. Louis, MO, USA).

Cell growth assay and focus formation assay. Cell growth rate was detected using the MTT assay. Briefly, cells were seeded in a 96-well plate at a density of 1.5x10⁴ cells. Cells were treated...
with baicalein (0, 10, 20, 30, 40, 50, 60, 80, 100 and 120 µM) (R&D Systems, Inc., Minneapolis, MN, USA) for 24 h. MTT assay was done in accordance with the manufacturer’s instructions. Each experiment was done in triplicate and data were expressed as mean ± SD. In focus formation assay, 100 cells were firstly seeded onto a plate and then cultured with (0, 40, 60 and 80 µM) baicalein for 2 weeks. The plates were fixed and then stained with 1% crystal violet. All experiments were performed in triplicate.

Construction of expression plasmids and transfection. The construction of the expression plasmids and their transfection was performed in accordance with the manufacturer’s instructions (3). First, the full-length pcDNA3.1 (Invitrogen, Carlsbad, CA, USA) MEK1 vector was made by cloning the full-length PCR product of MEK1 with KOD® DNA polymerase (Toyobo Co., Ltd., Osaka, Japan). Then we measured the plasmid sequences to confirm the sequences of the plasmid. In transient transfection experiments, cells were plated at a density of 2x10⁶ cells/well in a 6-well plate, 24 h later they were transfected with 4.0 µg pcDNA3.1(+)-MEK1 vector via Lipofectamine 2000 (Invitrogen) in accordance with the manufacturer’s protocol. The 4.0 µg pcDNA3.1(+) empty vector was used as a negative control.

Cell invasion and migration assay. The migration and invasion ability of cells were detected by Transwell assays. Migration and invasion assay was performed with 24-well Transwell chambers coated with Matrigel or not (Becton-Dickinson, Billerica, MA, USA) as previously described. For the migration assay, cells were seeded into inner well and cultured in medium with 0, 10, 20 and 30 µM baicalein. After 16 h, cells on the bottom side of the inner well were fixed in alcohol, fixed in crystal violet and counted. Inversion assay was performed with 24-well Transwell chamber coated with Matrigel as previously described (3).

Western blotting assay. After treated with U0126 or baicalein, 2x10⁶ cells were added into 200 µl of lysis buffer (Fermentas, Waltham, MA, USA). By 10% SDS-polyacrylamide gel electrophoresis, the proteins (60 µg) were separated and blotted onto PVDF membranes. To block non-specific binding, the PVDF membranes were subsequently blocked in Tris-buffered saline with Tween-20 (TBST) containing 5% defatted milk buffer for 1 h at 37°C and were then incubated with antibodies against ERK, p-ERK, MMP-2, MMP-9 or β-actin overnight in defatted milk 5% in TBST at 4°C. The membranes were incubated with a horseradish peroxidase goat anti-rabbit or anti-mouse IgG antibody at room temperature for 1 h. The pictures were examined with an enhanced chemiluminescence kit (ECL Plus; Amersham, Freiburg, Germany) according to the manufacturer’s instructions and captured by autoradiography. The relative photographic density was quantified with ImageJ software (GE Healthcare, Buckinghamshire, UK) and expressed as arbitrary units (a.u.).

Tumor xenograft experiments and animals. To assess the anti-proliferation of baicalein in vivo, Balb/c athymic nude mice (4- to 6-week-old) were obtained from Shanghai SLAC Laboratory Animal, Co., Ltd., (Shanghai, China). Each mouse was subcutaneously implanted with 1x10⁶ viable DLD1 cells in the right flank area. One group was treated with vehicle dimethyl sulfoxide (DMSO), the other group was treated with baicalein (20 mg/kg/day) for 3 weeks. At day 21, all the mice were sacrificed and the tumor tissues were removed and weighed. The experimental protocols used herein were evaluated and approved by the Animal Care and Use Committee of the Medical School of Xi'an Jiaotong University.

Results

Baicalein inhibits the proliferation of HT29 and DLD1 cells. The inhibition effects of baicalein on the proliferation of CRC cells at different concentrations (0 to 120 µM) for 24 h are shown in Fig. 1B. At concentrations >40 µM, the anti-proliferation effect of baicalein on HT29 and DLD1 cells was significant so we chose concentrations <40 µM for all the following experiments.

To determine the effect of baicalein on apoptosis in detail, HT29 cells were incubated for 48 h with baicalein (40, 60 or 80 µM) and then analysed by flow cytometry. As shown in Fig. 1C, the effect of baicalein on apoptosis of HT29 cells was concentration-dependent. After treatment for 48 h, both early and late apoptotic cells increased significantly in the HT29 cells treated with baicalein. Baicalein reduced the number of colony HT29 cells in vitro significantly (Fig. 1D).

Baicalein inhibits the metastasis of HT29 and DLD1 cells. Fig. 2 shows the inhibition effect of baicalein (0, 10, 20 and 30 µM) on cell migration (16 h) and invasion (24 h) in HT29 and DLD1 cells, respectively. We found that inhibition effect of baicalein on the migration (Fig. 2A and B) and invasion (Fig. 2C and D) of HT29 and DLD1 cells was concentration-dependent.

Baicalein inhibits the expression of MMP-2/-9. We analysed the effect of baicalein on the expression of MMP-2/-9 in HT29 and DLD1 at various concentrations. Cells were treated with baicalein (0, 10, 20 and 30 µM) for 24 h and then analysed by western blotting. Fig. 3A and B show the inhibited effect of baicalein on the expression of MMP-2/-9 in HT-29 cells. The effect is concentration-dependent. Fig. 3C and D show the effect of baicalein on the expression of MMP-2/-9 in DLD1 cells.

The ERK signaling pathway is involved in the anti-metastatic mechanism of baicalein in CRC. ERK signaling pathway plays an vital role in the metastasis of cancer cells by regulating MMP-2/-9 (3); hence, we detected the impact of baicalein on the activity of ERK signaling pathway in CRC cells. The results of the western blotting showed that baicalein reduced the phosphorylation level of ERK1/2 in DLD1 cells treated with baicalein (Fig. 4A and B). To further check the role of ERK signaling pathway in the anti-metastatic effect of baicalein, we upregulated the activity of ERK singnaling pathway in DLD1 cells via transfecting the plasmid [pcDNA3.1 (+)-MEK1] expressing human MEK1 and found that the anti-invasion effect of baicalein was reversed by the high expression of MEK1 (Fig. 4C). The positive clones were selected by G418. DLD1 cells were
separated into 3 groups: no treatment as the control group ‘C’, transfected with an empty vector pcDNA3.1(+) as the negative control group ‘N’, transfected with a pcDNA3.1(+)–MEK1 as the positive group ‘M’.

Figure 1. Chemical structure of baicalein and the anti-proliferation effect of baicalein. (A) Chemical structure of baicalein. (B) The cell viability was measured by MTT assay. (C) Baicalein inhibited the colony formation ability of CRC cells. (D) Baicalein induced the apoptosis of CRC cells. Values are represented as means ± standard deviation (SD) of three independent experiments performed in triplicate. *P<0.05 and **P<0.01 compared with the control group, respectively.

Figure 2. Baicalein inhibits the migration and invasion of CRC cells. (A) HT29 and DLD1 cells were pretreated with 0, 10, 20 and 30 µM baicalein for 24 h. Then seeded onto the upper wells, FBS (10%) was added to the bottom chambers for 16 h to induce cell migration. After 16 h, cells on the bottom side of the filter were fixed, stained and counted. (B) The percentage of migration rate was expressed as a percentage of control (0 µM). (C) HT29 and DLD1 cells were pretreated with 0, 10, 20 and 30 µM baicalein for 24 h. Then seeded onto the upper wells, FBS (10%) was added to the bottom chambers for 24 h to induce cell invasion. After 24 h, cells on the bottom side of the filter were fixed, stained and counted. (D) The percentage of invasive rate was expressed as a percentage of control (0 µM). Values are presented as means ± SD of three independent experiments performed in triplicate. *P<0.05 compared with the control group, respectively.
The inhibition rate was ~61.3 and 39.3% after 24 h of treatment with 0 or 30 µM of baicalein (Fig. 4D). Baicalein could also inhibit the phosphorylation level of ERK in the HT29 cells (data not shown). Our results also show that the combined treatment with the baicalein and U0126 (an ERK inhibitor) reduced both the MMP-2/-9 protein expres
sion (Fig. 5A and B) and the cell invasion significantly (Fig. 5C and D).

**Baicalein inhibits the growth of CRC tumors in vivo.** The inhibition effect of baicalein on DLD1 xenograft growth is shown in Fig. 6A. We found that baicalein could inhibit CRC growth significantly. At day 21, all the mice were sacrificed and the tumor tissues were removed and weighed. Compared to the control group, baicalein reduced the volume of solid tumors significantly (Fig. 6B). We also found that baicalein significantly inhibited phosphorylation level of ERK and the expression of MMP-2/-9 in vivo. The inhibition effect of baicalein on the phosphorylation level of ERK is shown in Fig. 6C and D, and on expression of MMP-2/-9 is shown Fig. 6E and F in vivo.

**Discussion**

Baicalein possesses antitumor effect in many tumors (17-19), but the reports on baicalein effect are rare in CRC. We found that baicalein significantly inhibited the migration and invasion of CRC cells by inhibiting MMP-2/-9 expression via inhibiting the ERK signaling pathway.

Our results showed that baicalein inhibited the proliferation and colony formation ability of CRC cells and the mechanism is correlated with apoptosis. Transwell chamber is generally used as an assay to detect the metastasis ability of cancer cells. In the study, we chose the concentration below cytotoxicity to detect the effect of baicalein on the migration of CRC cells. We found that baicalein could inhibit the proliferation of hepatocellular carcinoma cells, glioma and breast cancer cells. As concentrations are below the cytotoxicity concentration, the inhibition effect is not correlated with the cytotoxicity of baicalein. To the best of our knowledge, we first show the effect of baicalein on DLD1 CRC cells which was used for research on proliferation, apoptosis and invasion of CRC (20-22). The inhibitory properties are probably related to the specific structural features of baicalein (as shown in Fig. 1A). It was reported that the hydroxylation of C5 and C7 in A-ring significantly improved the anticancer activity of flavonoids over that of 5-fluorouracil (23). Moreover, it has been demonstrated that the hydroxyl substitutions in the A-ring (C7) of baicalein are crucial for its anti-metastatic effects against human hematoma cells (24).

To date, metastasis of CRC causes thousands of deaths every year worldwide (25). Degradation of the ECM of lymph or blood vessels exert important roles in the metastasis of
cancers via allowing the invasion of cancer cells into the circulation system and invade distant organs or tissues (26). MMPs, especially MMP-2/-9, play important roles in degrading ECM (27). Baicalein shows inhibition effect on the expression of MMP-2/-9 in hepatocellular carcinoma and glioma (3,16). Our results showed the inhibition effect of baicalein on MMP-2/-9 expression in CRC cells. The results suggest that MMP-2/-9 play important roles in the anti-metastatic effect of baicalein in CRC cells.

Multiple signaling cascades play important roles in the synthesis of MMPs, especially the ERK signaling pathway (19,28,29). ERK signaling pathway plays an important role in tumor invasion via promoting the degradation of ECM proteins (30,31). Studies have shown that the inhibition of ERK phosphorylation reduces the expression of MMP-2/-9 in the CRC cells (32,33). To investigate the related mechanism(s) of the inhibition effect of baicalein on CRC metastasis, we detected the activity of ERK signaling pathway in CRC cells. The results proved that baicalein reduced the activity of ERK signaling pathway significantly. Upregulation of ERK signaling pathway activity abolished the baicalein's antimetastatic effect. The results was similar to a previous study, in which baicalein inhibited the expression and activity of MMP-2 and MMP-9 via inhibiting the activity of ERK signaling pathway (3).

Our results also showed that combined treatment with baicalein and U0126 (an ERK inhibitor) reduced both the MMP-2/-9 protein expression (Fig. 5A and B) and the cell invasion significantly (Fig. 5C and D) suggesting that baicalein directly downregulated the ERK signaling pathway, which were similar to previous studies conducted in other types of carcinomas (3). Finally, our results demonstrated that baicalein can inhibit the growth of CRC xenografts in vivo.

In conclusion, the present study demonstrated the anti-metastatic effect of baicalein on CRC. Furthermore, ERK signaling pathway plays a vitally important role in the anti-metastatic effect of baicalein on CRC cells by regulating MMP-2/-9. These findings revealed that baicalein may represent a new potential anti-metastatic therapy for CRC.

References


